

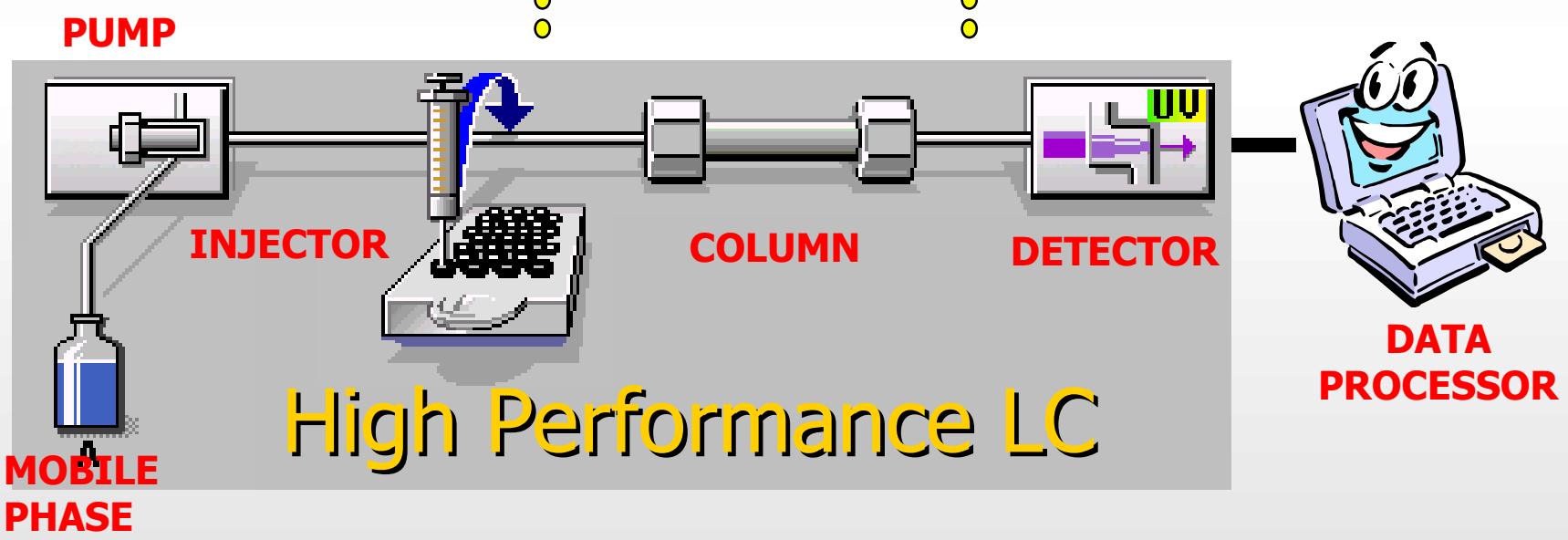
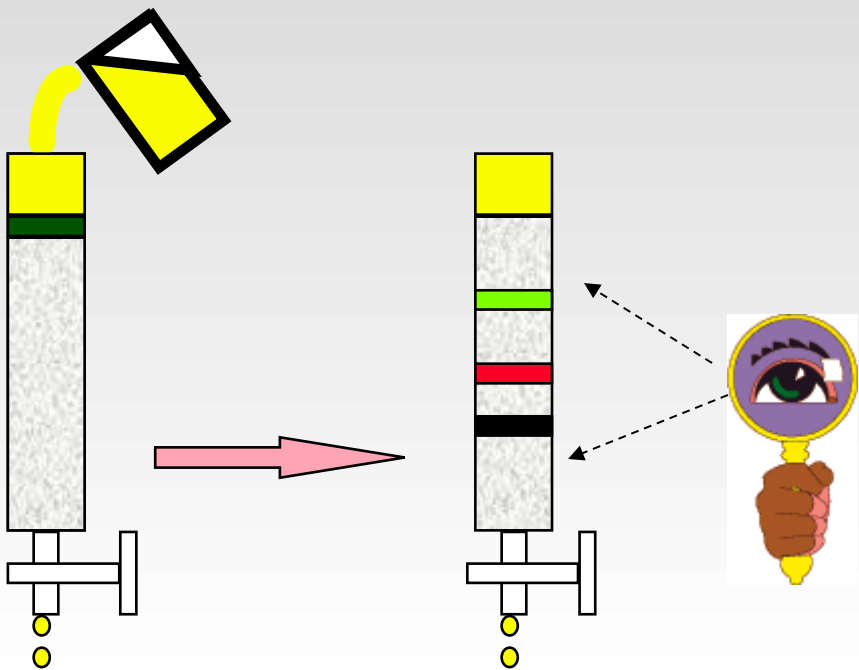
# Introduction to HPLC



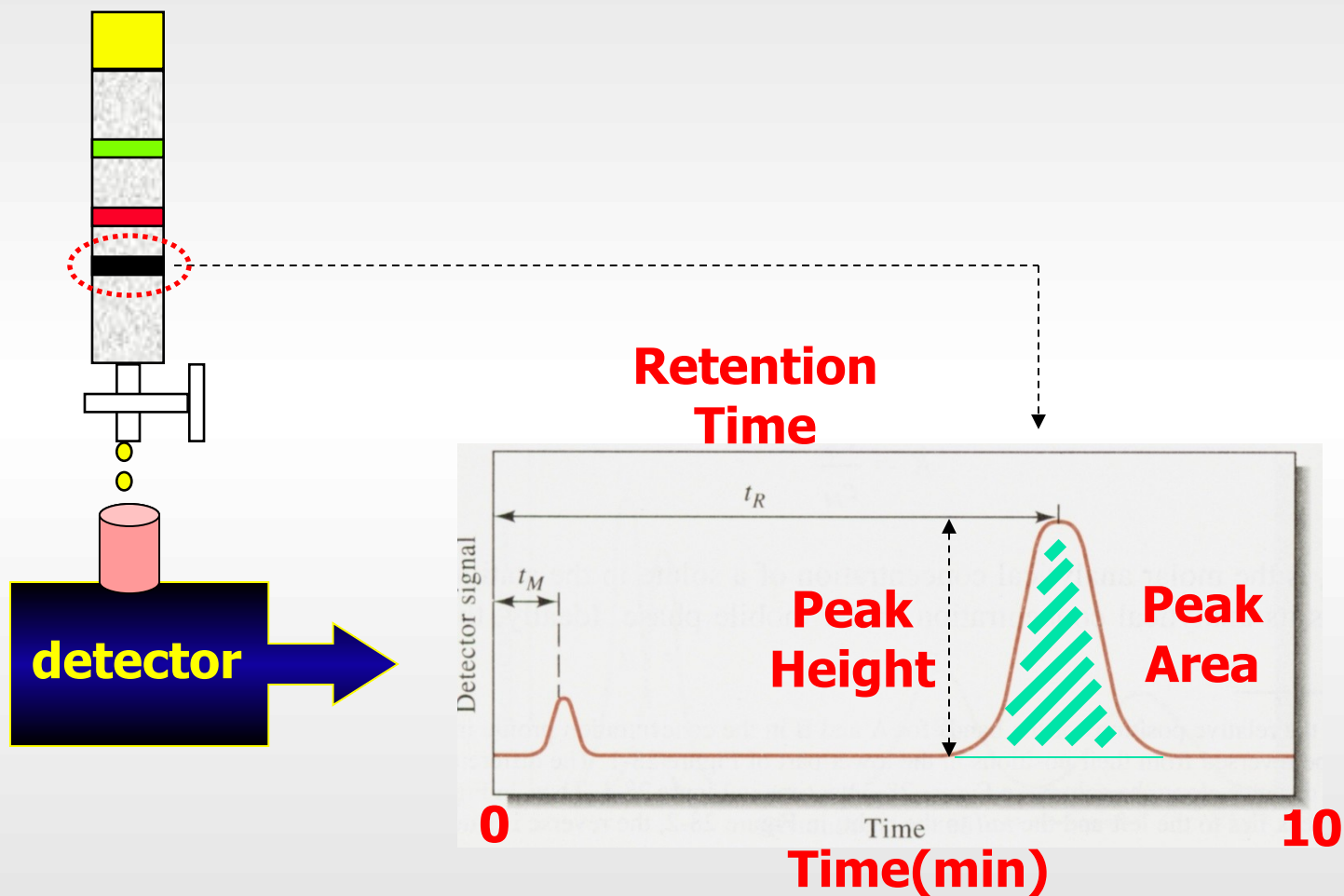
**SPC-CSC**

# Applications for HPLC

- Pharmaceuticals
  - Antibiotics
  - Vitamins
  - Antipyretic & Analgesic drugs
- Environmental
  - Inorganic ions
  - Pesticides
- Polymers
  - Antioxidants
  - Plasticizers
- Food
  - Preservatives
  - Vitamins
  - Sugars
  - Organic acids
- Medical
  - Amino acids
  - Drugs
  - Metabolites



# Chromatographic Data



# Modes of HPLC

- **Normal Phase mode**
- **Reverse Phase mode**
- Reverse Phase Ion Pairing mode
- Ion Exchange mode
- SEC mode (GPC / GFC)
- Chiral separation mode

# Normal Phase Mode

- First technique used
  - $\text{CaCO}_3$  in Separation Column
  - Petroleum ether as Eluting Solvent
- We define this combination as

## *Normal Phase mode*

Column : polar property

Solvent : non-polar property

# Reverse Phase Mode

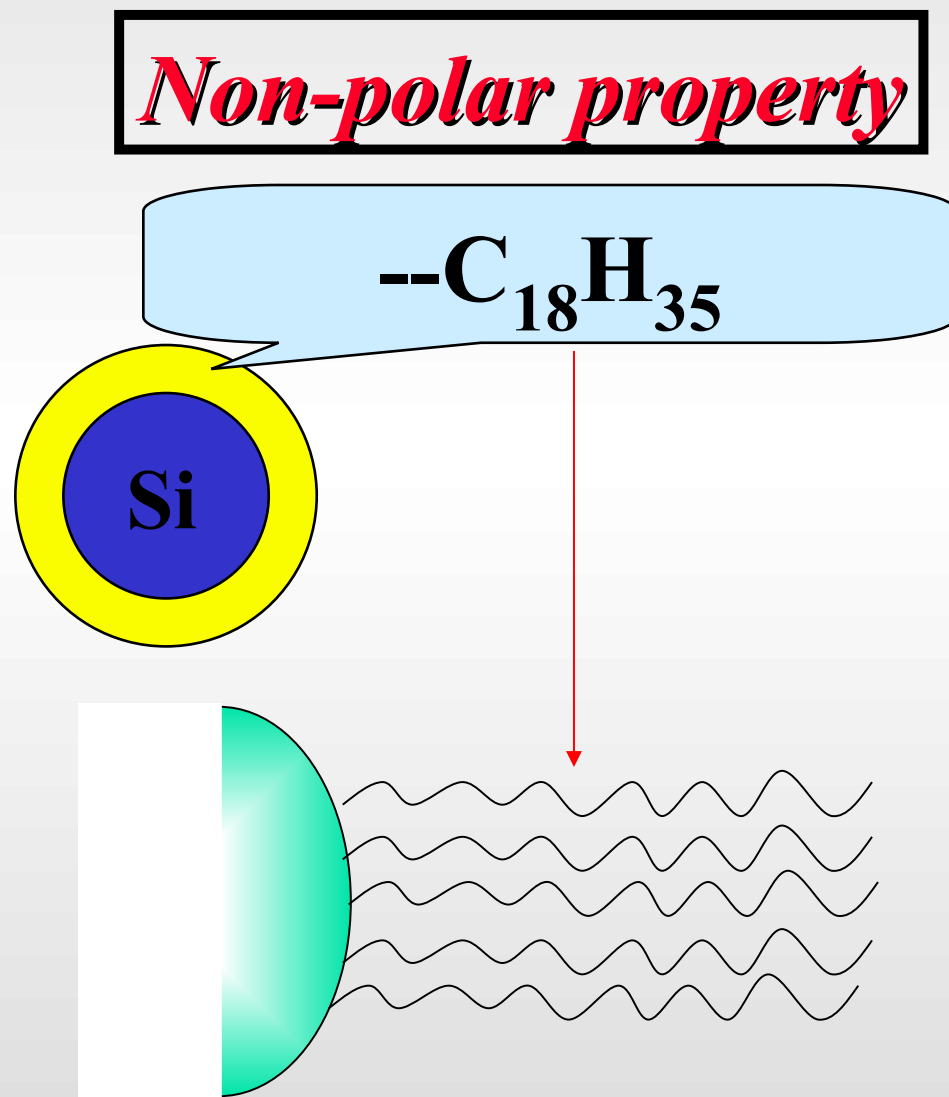
**Column : Non-polar property**

**Solvent : Polar property**

***water /methanol / acetonitrile***

# Reverse Phase HPLC Columns

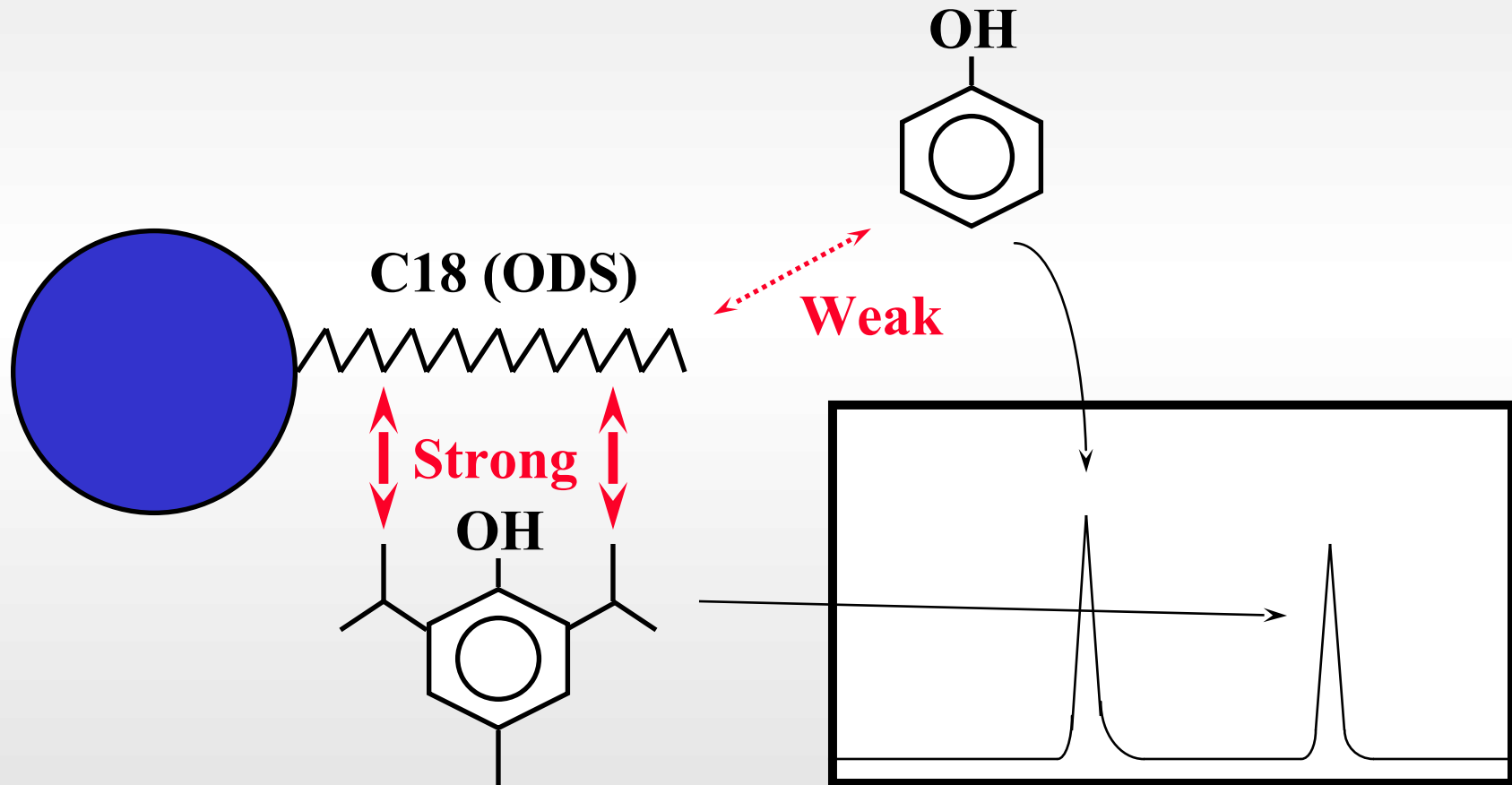
- C18 type
- C8 (octyl) type
- C4 (butyl) type
- Phenyl type
- TMS type
- Cyano type



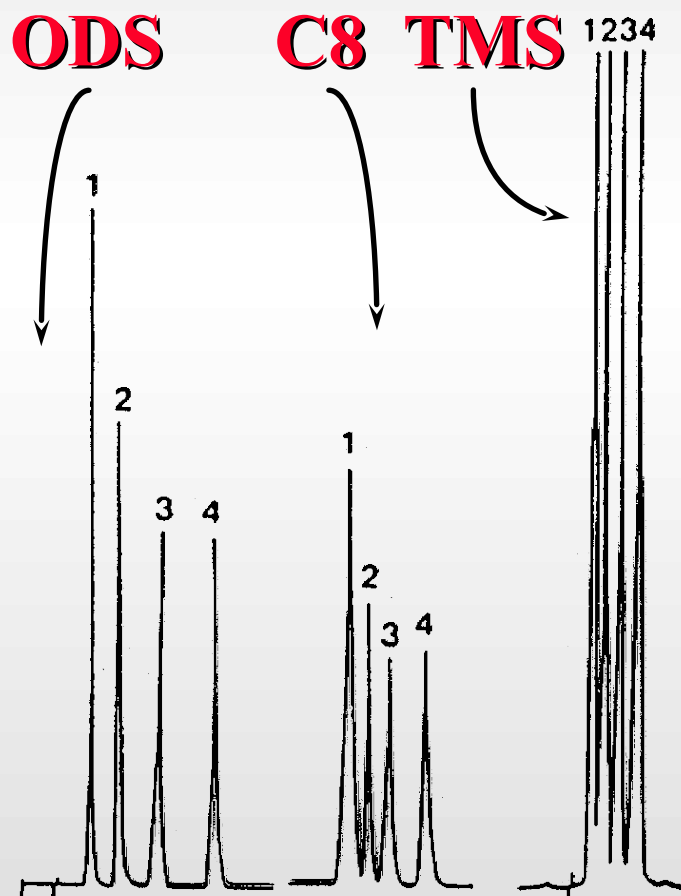


# Principle of separation

“like dissolves like”



# Effect of stationary phase



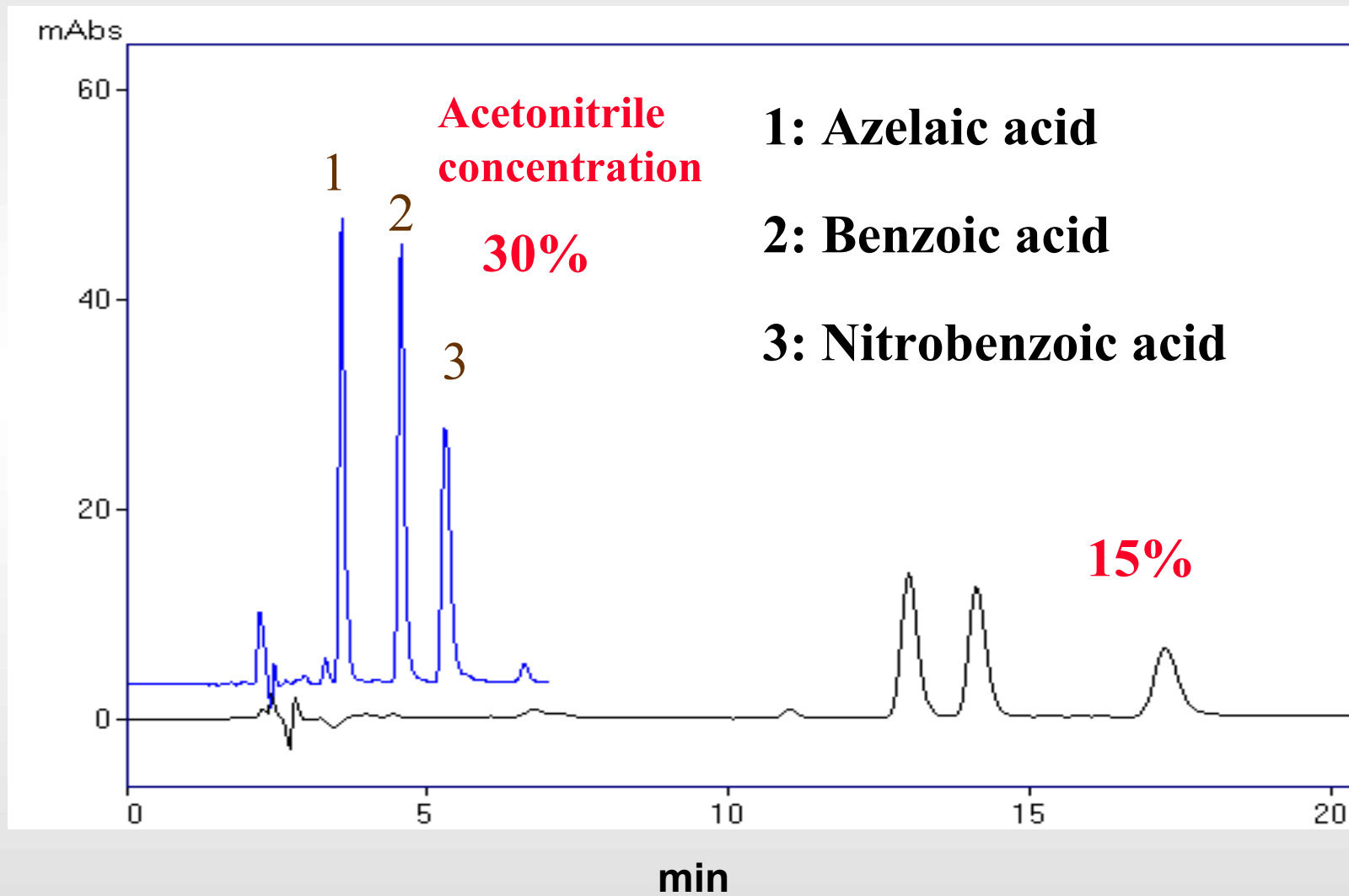
## ◆ Analytical Conditions

- ❖ Column : Shim-pack CLC-ODS
- ❖ Mobile phase : MeOH : H<sub>2</sub>O = 7 : 3
- ❖ Flow rate : 1.0 mL/min
- ❖ Temperature : 40 C
- ❖ Injection volume : 10 uL
- ❖ Detection : UV-254 nm

## ◆ Peaks

1. Methyl benzoate
2. Ethyl benzoate
3. n-Propyl benzoate
4. n- Butyl benzoate

# Effect of Mobile Phase Composition



# Normal vs Reverse Phase

- **Normal Phase**

- good separation for stereo isomer  
(Vitamin E etc..)
- variable retention time

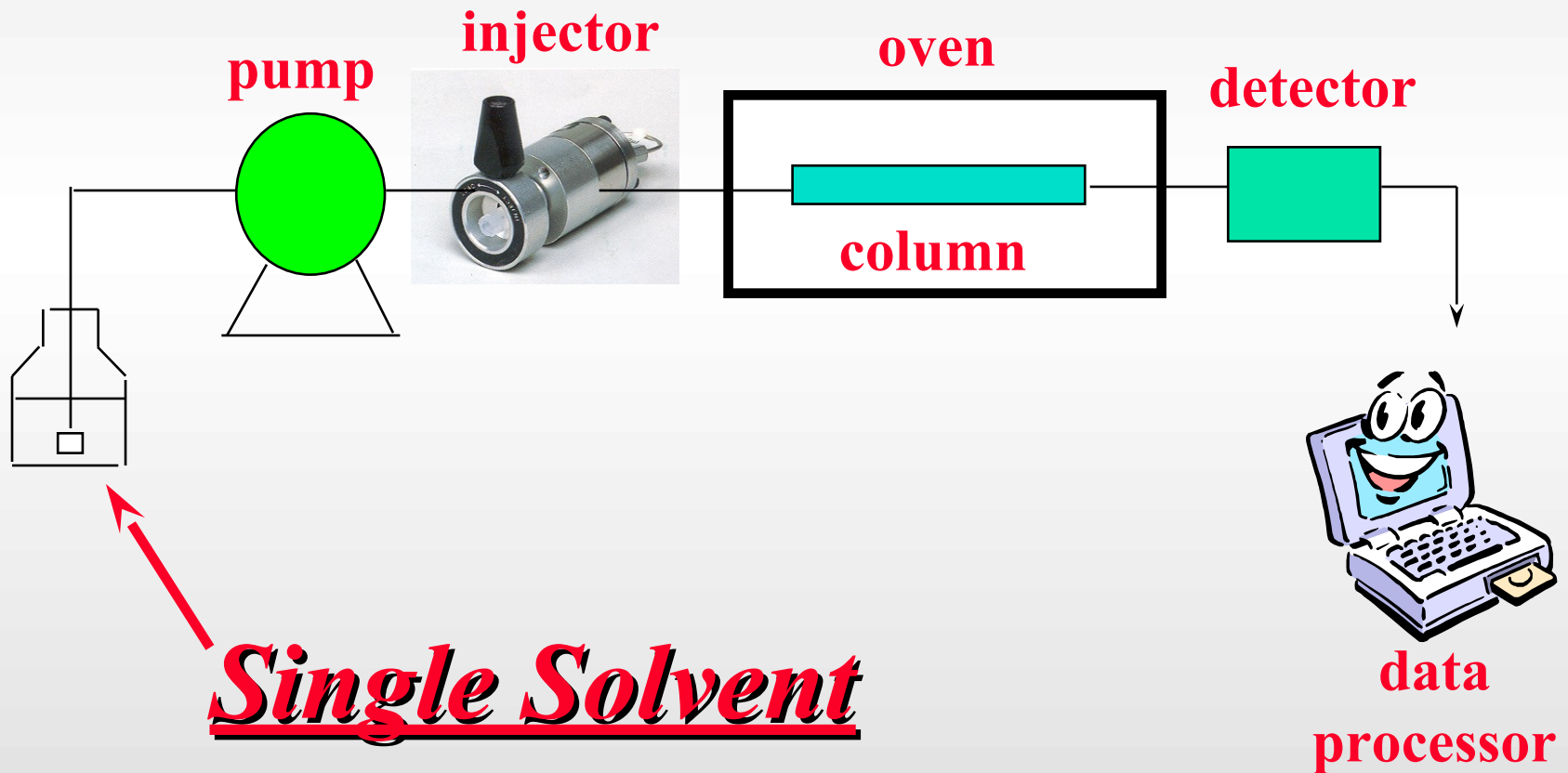
- **Reverse Phase**

- good repeatable retention time
- rugged stationary phase

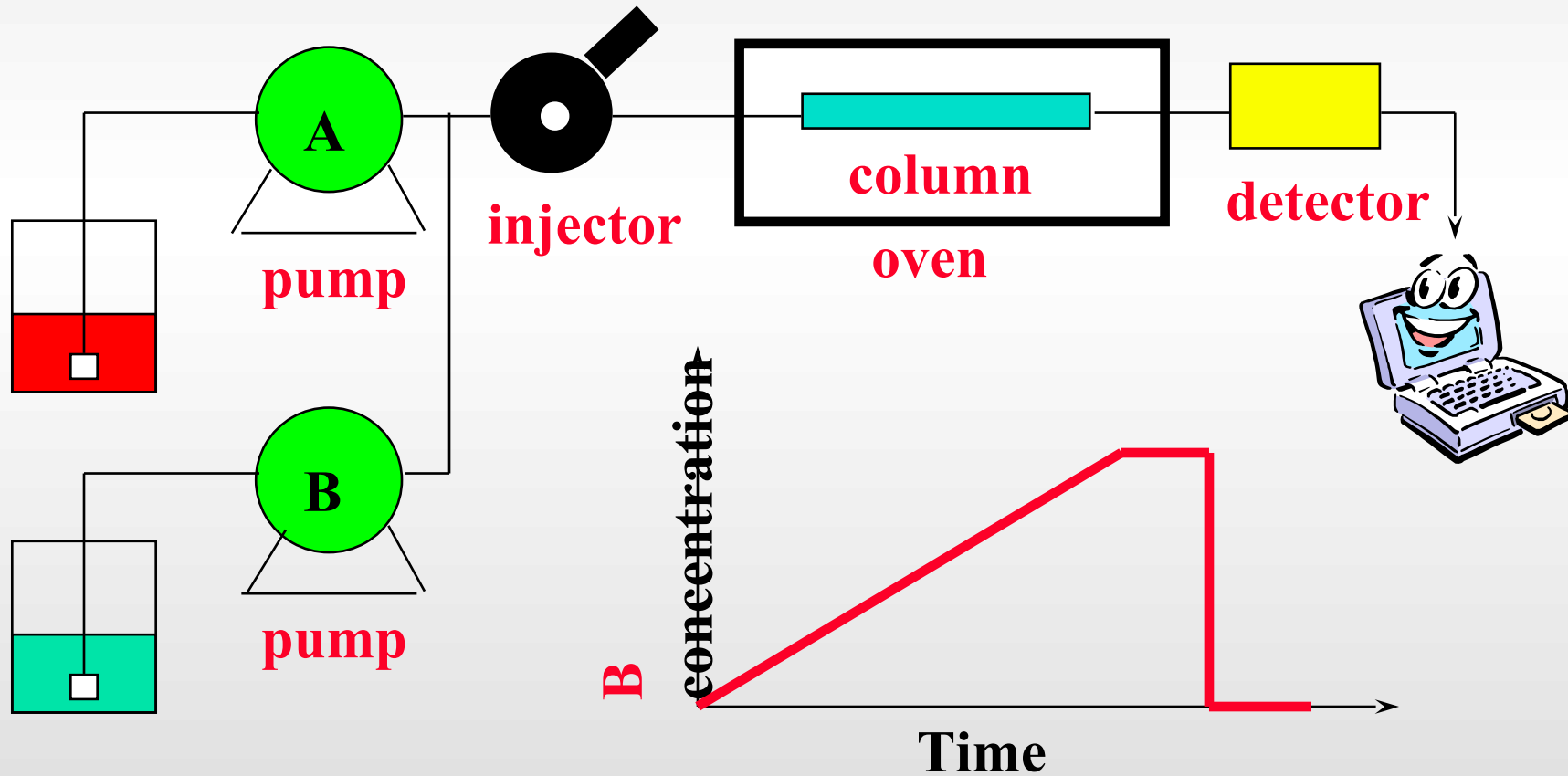
# HPLC System

- Isocratic elution system
  - Single solvent of constant composition
- Gradient elution system
  - Multiple solvents of variable composition
    - High pressure gradient system
    - Low pressure gradient system

# Isocratic Elution System



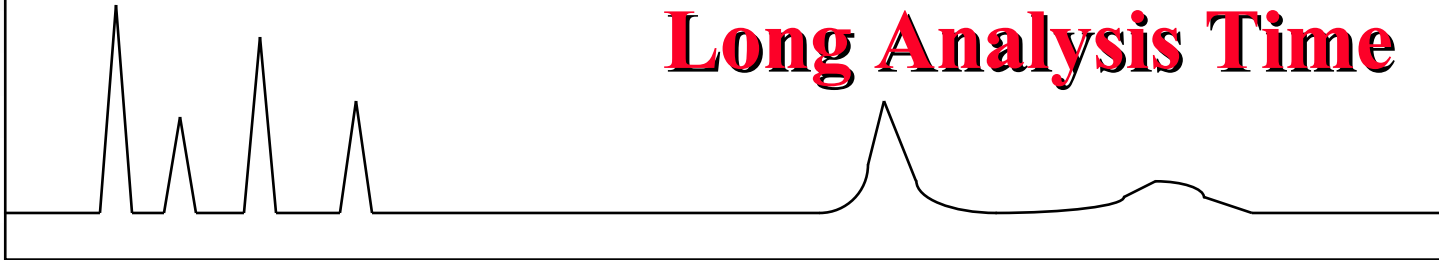
# Gradient Elution System



# Isocratic Elution Mode

**MeOH / H<sub>2</sub>O = 6 / 4**

**Long Analysis Time**



**Bad Separation**

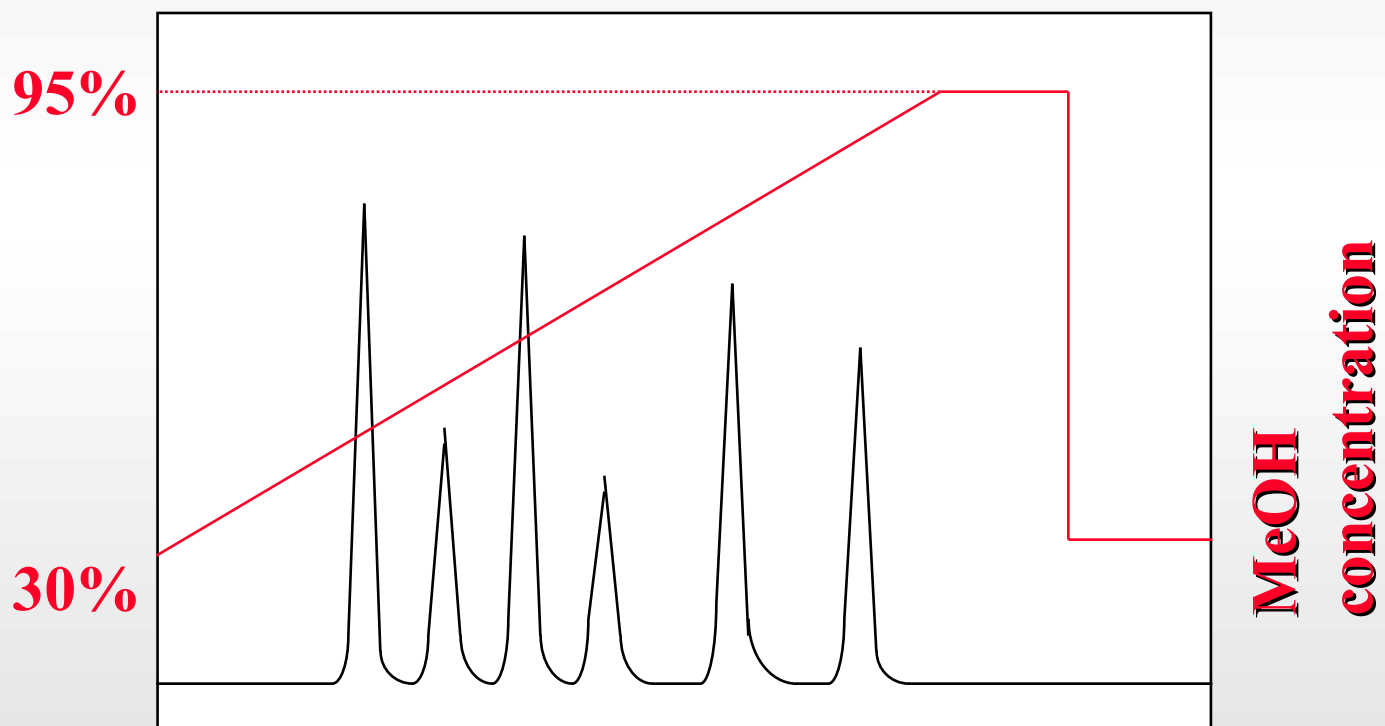
**MeOH / H<sub>2</sub>O = 8 / 2**



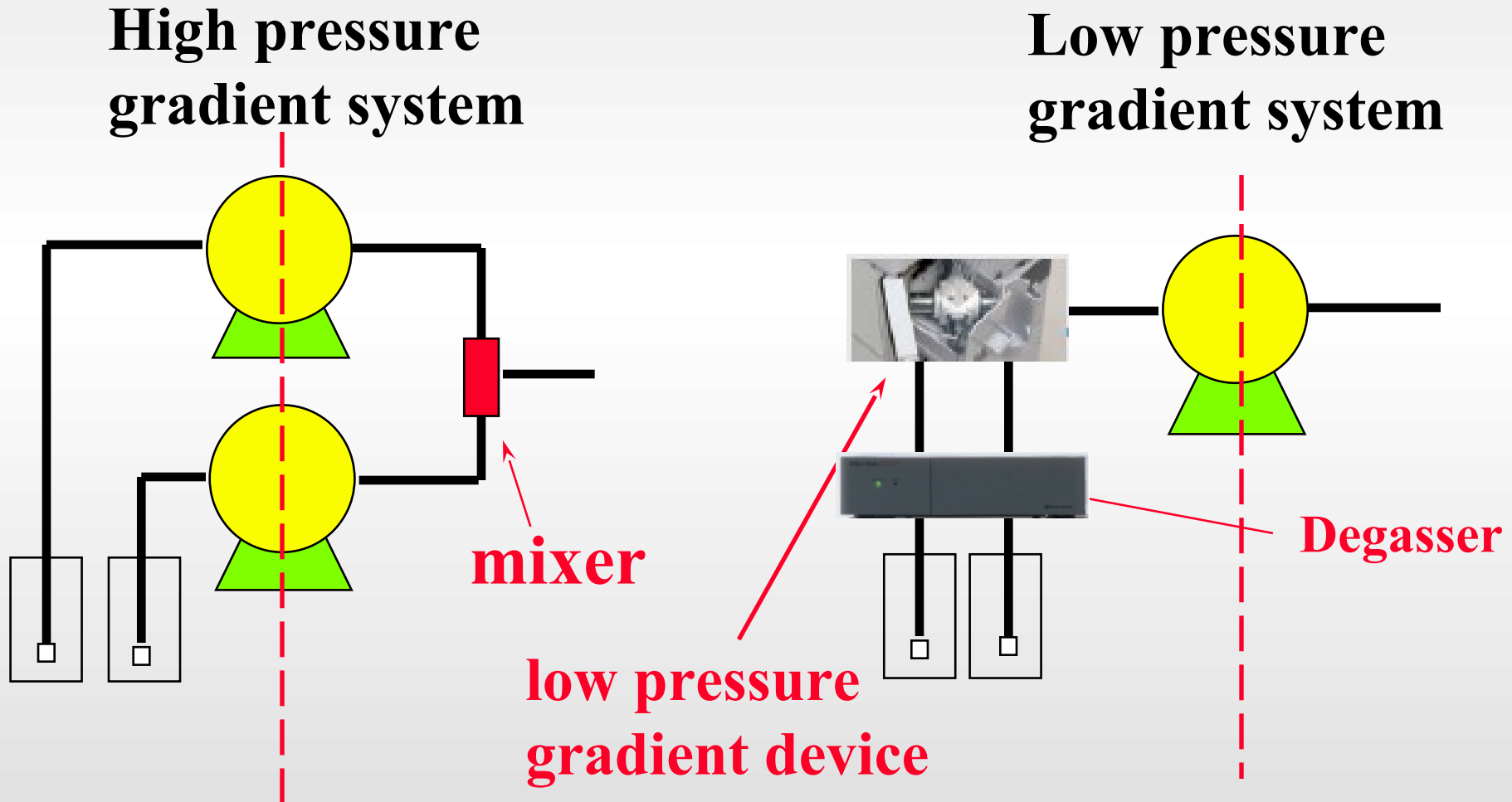
( column : ODS type )



# Gradient Elution Mode



# High/Low pressure gradient system



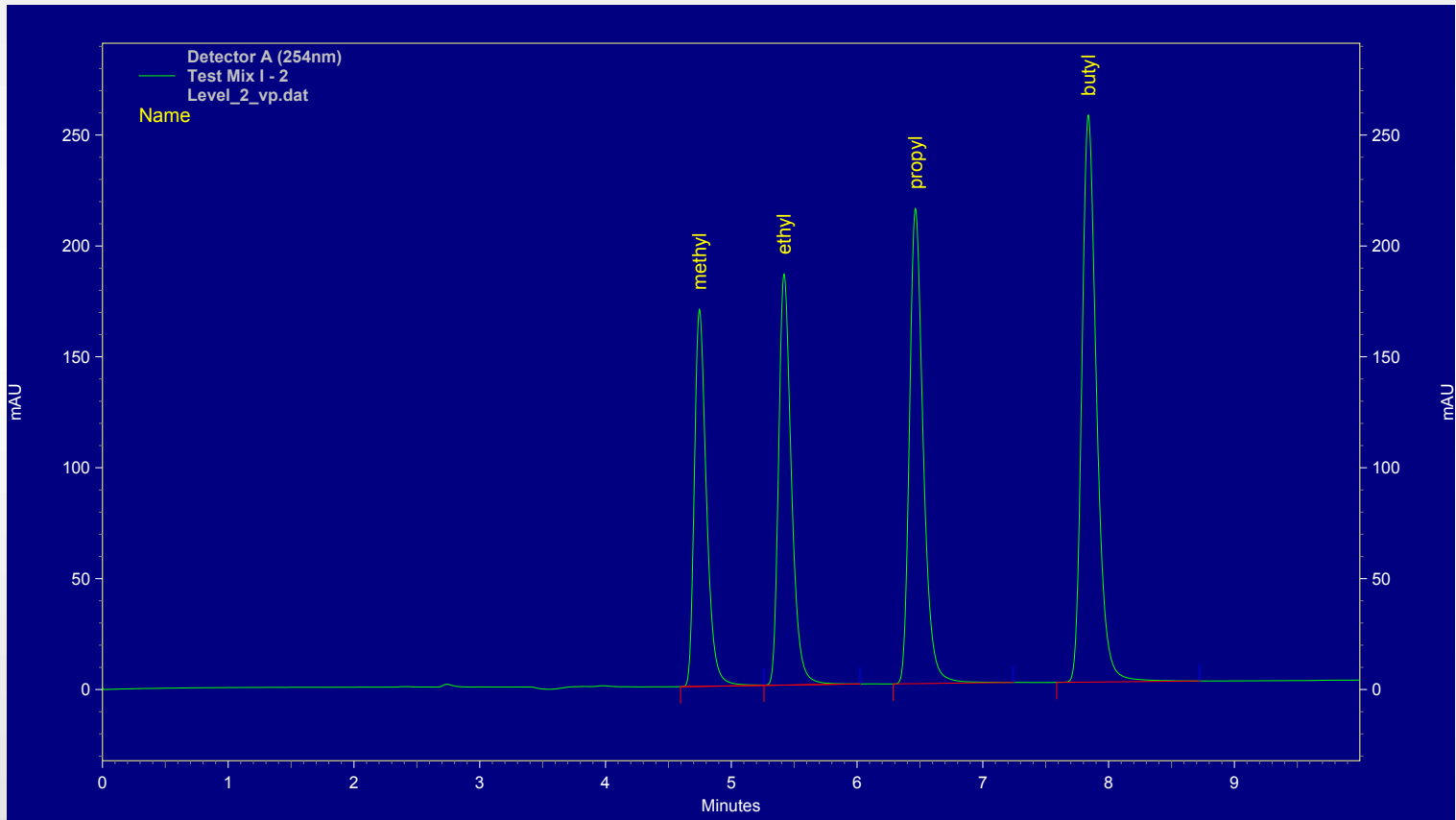
# High/Low pressure gradient system

- High pressure gradient system
  - excellent gradient accuracy
  - complicated system (more than two pumps)
- Low pressure gradient system
  - simple system
  - degasser is required

# QUALITATIVE/QUANTITATIVE ANALYSIS

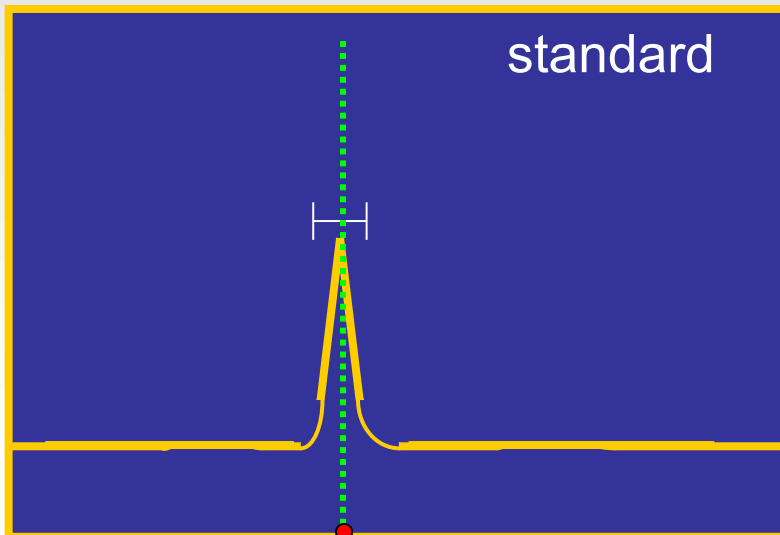
# QUALITATIVE ANALYSIS

Detector Response

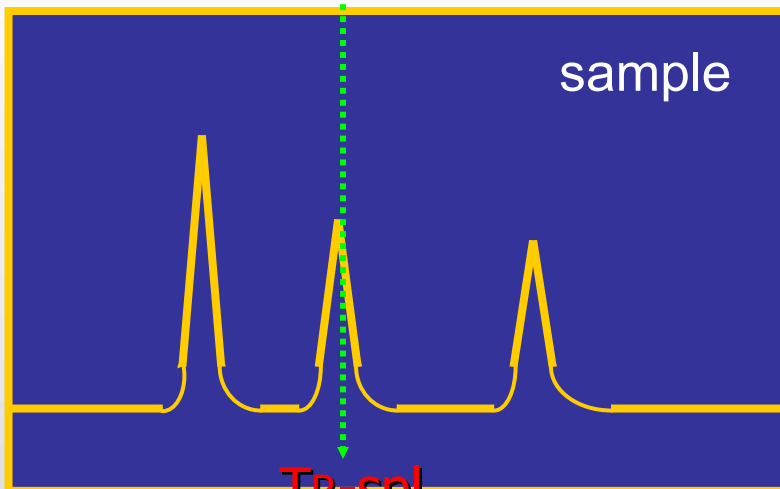


Retention Time

# QUALITATIVE ANALYSIS



TR-std



TR-spl

**Basic Question:**

**Does the sample  
have a peak with a  
TR-spl within  $\pm X\%$   
of the TR-std?**

# QUALITATIVE ANALYSIS

- ⊕ Identification of individual components in the sample
- ⊕ STANDARDS of known composition are needed
- ⊕  $T_R$  (retention time) is the qualitative data
- ⊕ Directly compare the  $T_R$  of the standard and the unknown



# QUANTITATIVE ANALYSIS

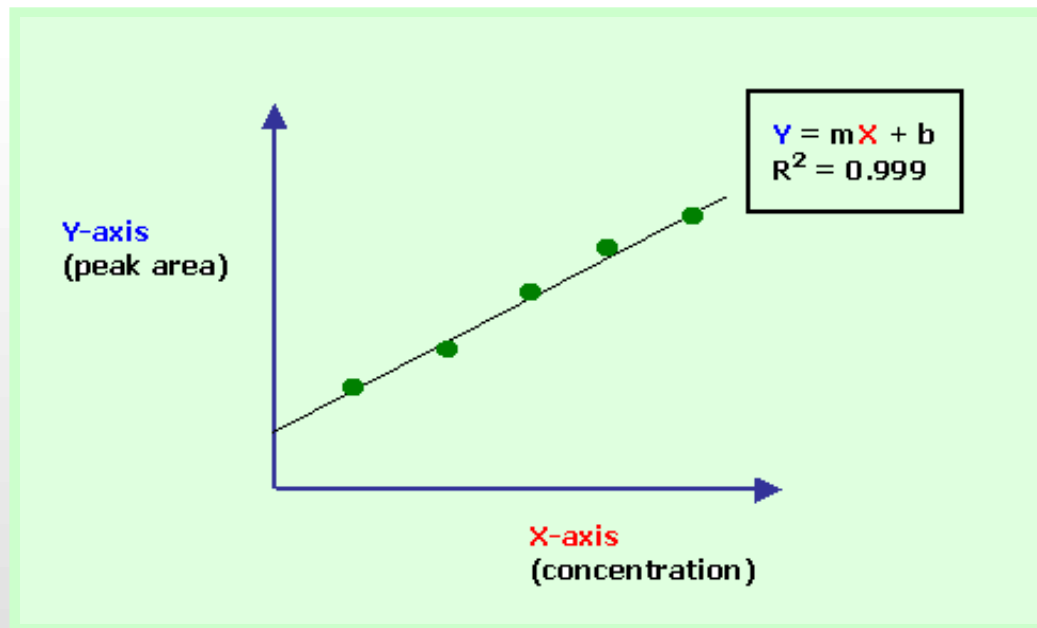
- ⊕ **Determination of the amount / concentration of individual components separated in the sample**
- ⊕ **Peak area or peak height is the quantitative data ( $\alpha$  concentration)**
- ⊕ **STANDARDS of known composition & concentrations are needed**





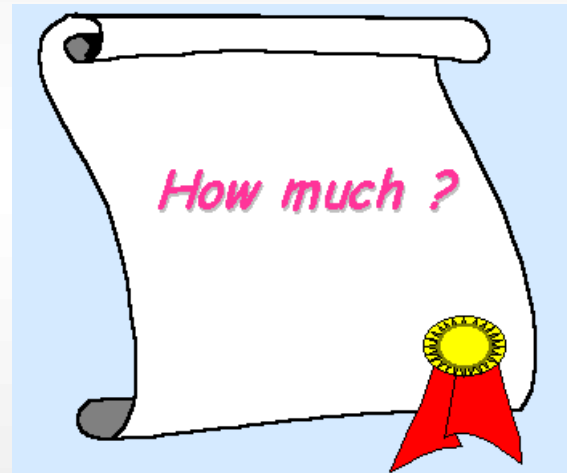
# QUANTITATIVE TERMS

- **Calibration/Standardization**: generation of a curve that shows the relationship between concentration and peak area/height (per component)
- **External/Internal standardization method**: two of the most popular calibration methods

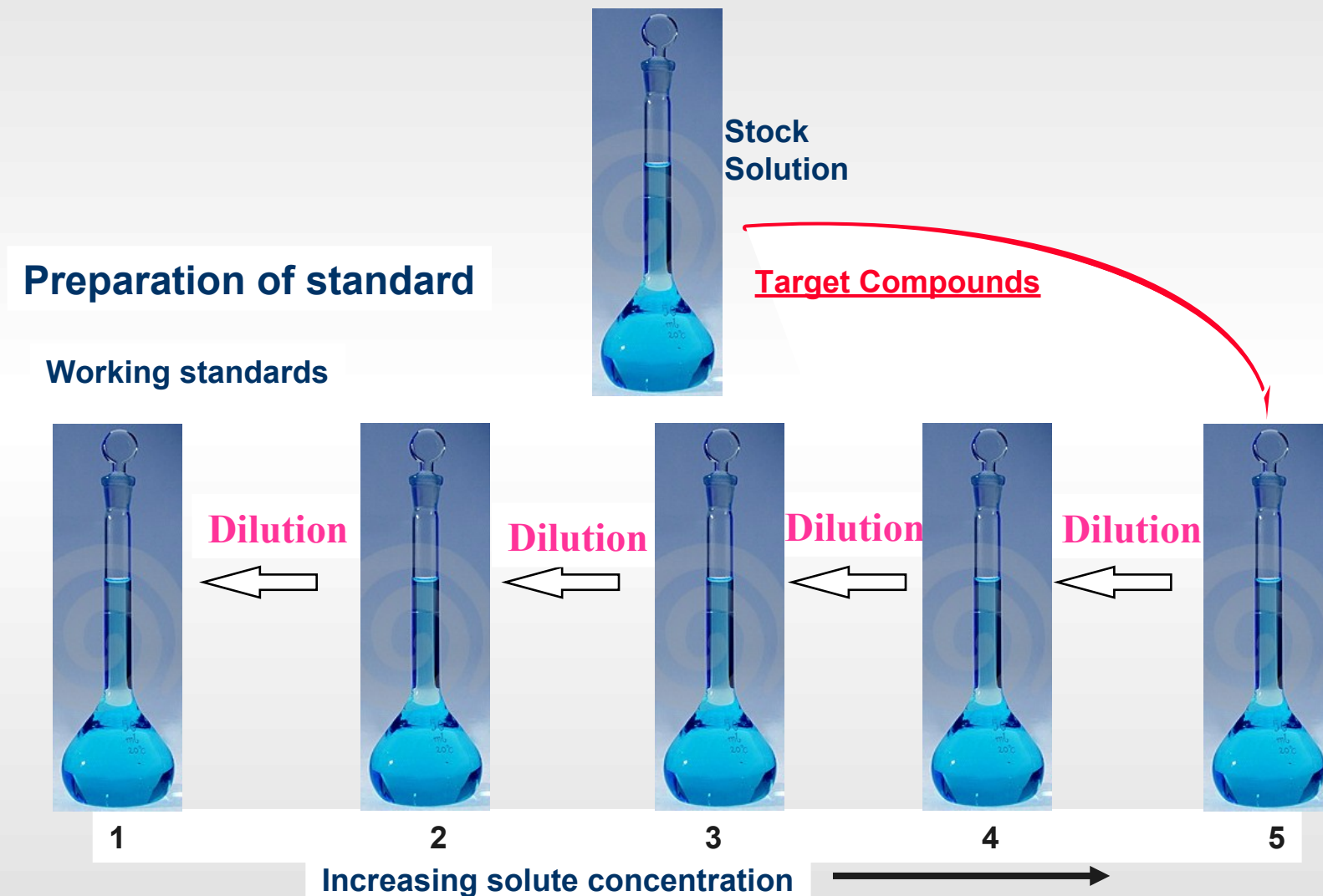


# QUANTITATIVE TERMS

- **External Standard Method**
- **Internal Standard Method**

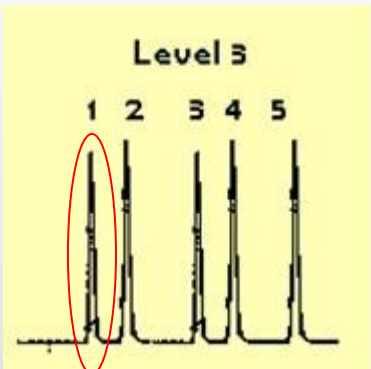
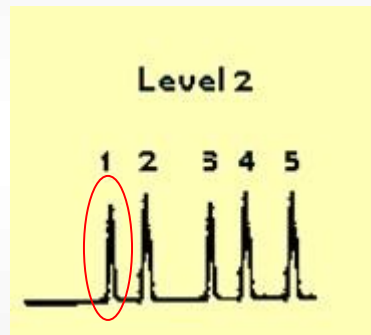
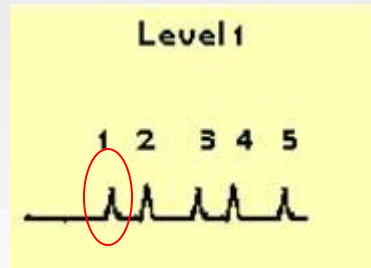


# EXTERNAL STANDARD METHOD

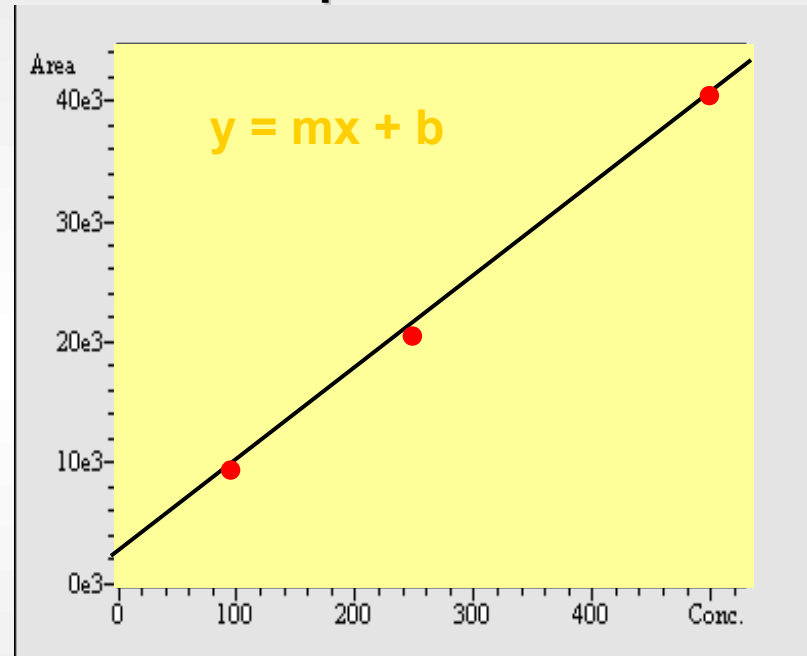


# EXTERNAL STANDARD METHOD

Increasing  
concentration



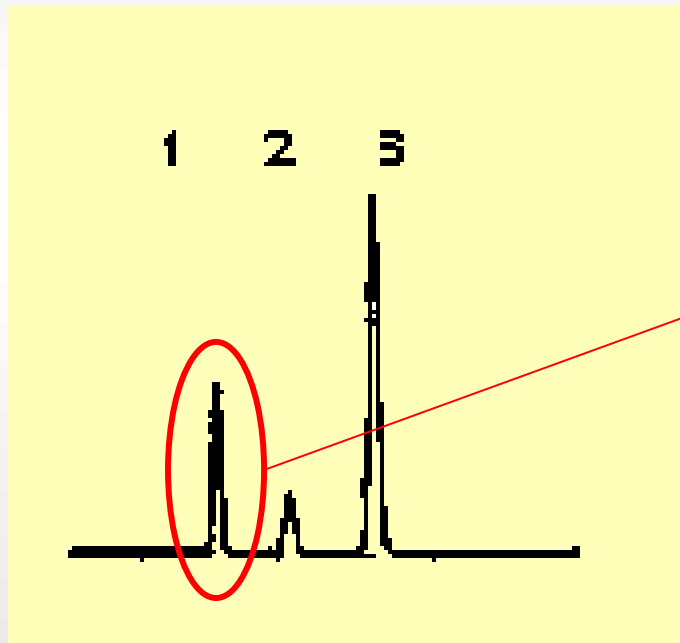
3-point calibration curve  
for peak # 1



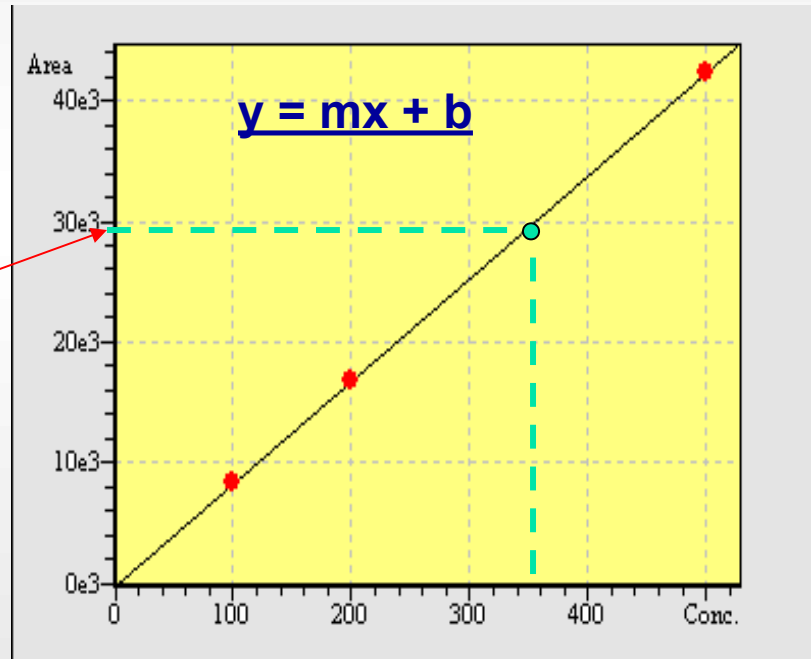
**Y = AREA or HEIGHT**  
**X = CONC.**

# EXTERNAL STANDARD METHOD

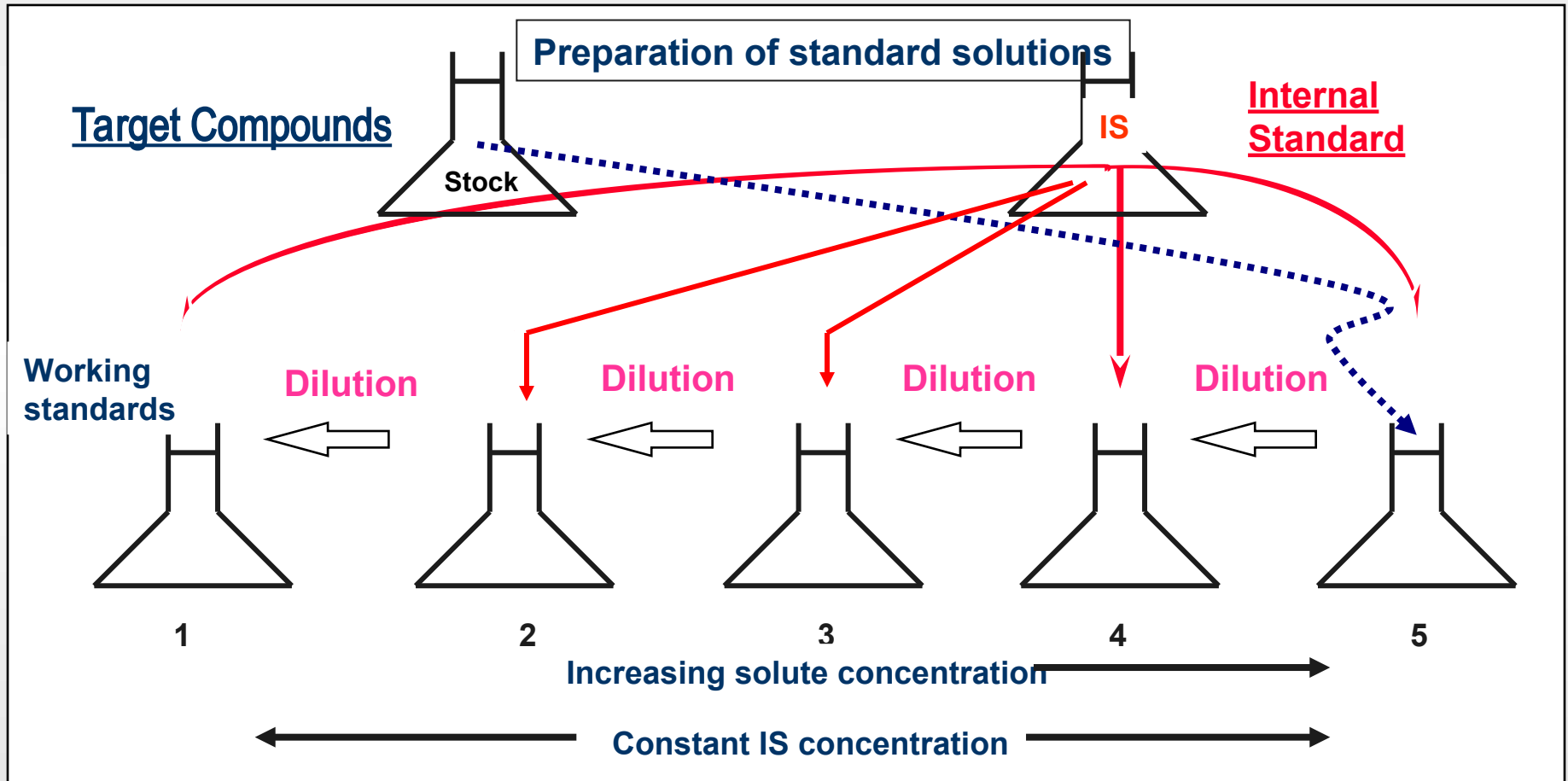
Chromatogram for sample



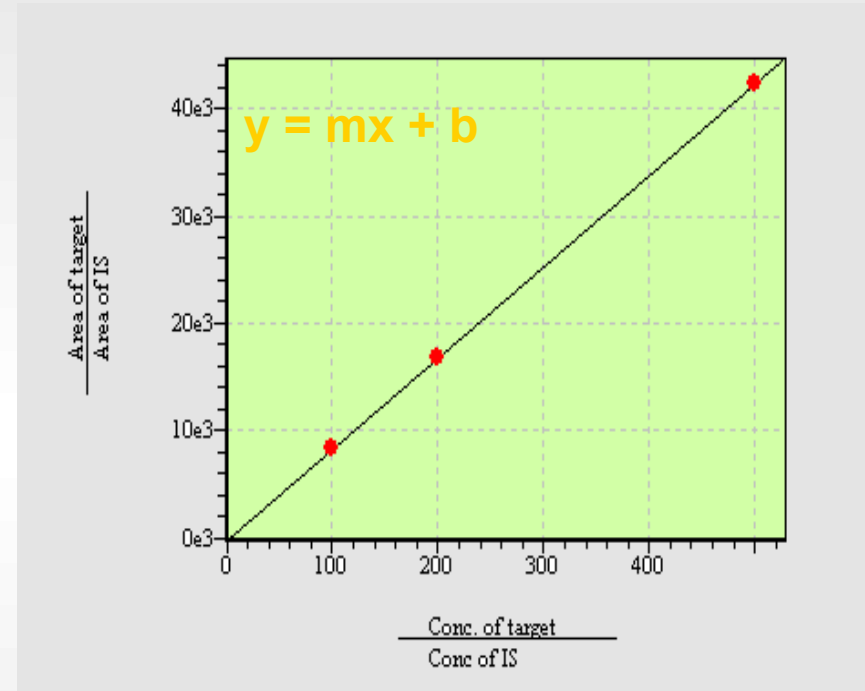
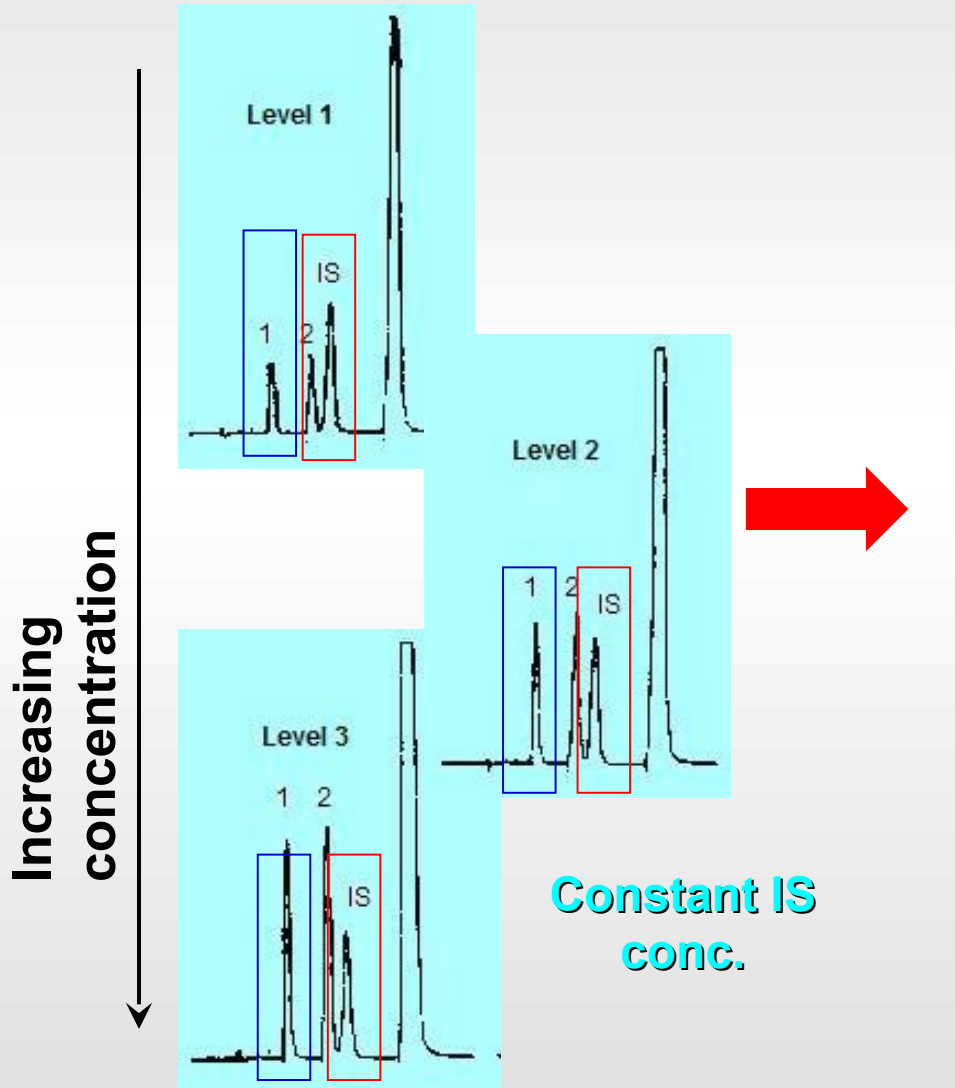
Calibration curve  
(for peak 1)



# INTERNAL STANDARD METHOD



# INTERNAL STANDARD METHOD

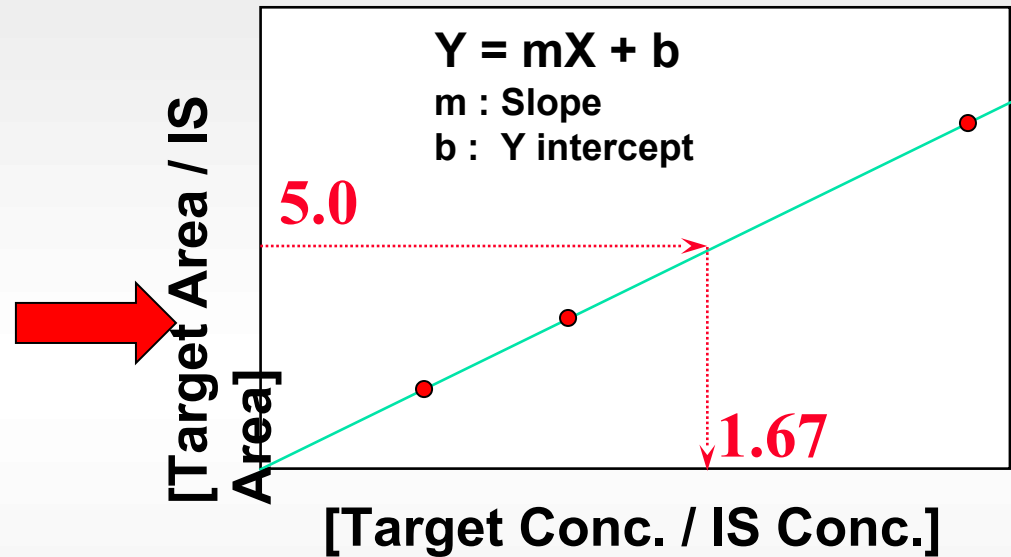
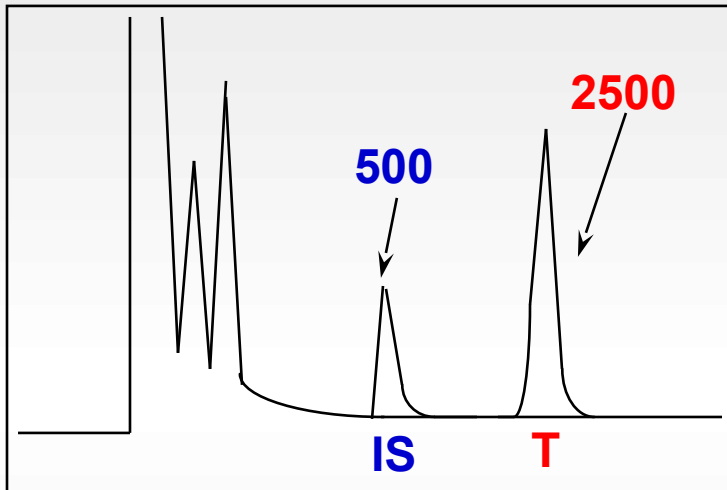


**Y = AREA/ HEIGHT RATIO  
(target/IS)**

**X = CONC. RATIO  
(target/IS)**

# INTERNAL STANDARD METHOD

## Calculation of Results



$$Y = mX + b$$

$$\frac{\text{Area T}}{\text{Area IS}} = m \cdot \frac{\text{Conc T}}{\text{Conc IS}} + b$$

$$X = \text{Target Conc.} / \text{IS Conc.}$$

$$1.67 = \text{Target Conc.} / 100 \text{ ppm}$$

$$\text{Target Conc.} = 167 \text{ ppm}$$



# INTERNAL STANDARD METHOD

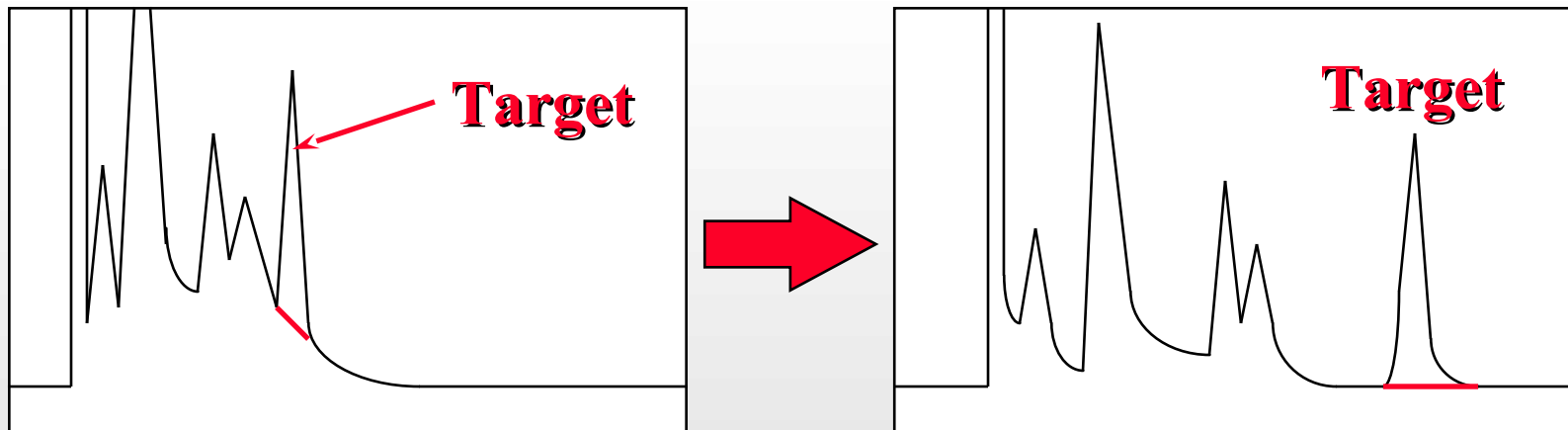
- Properties of internal standard
  1. Must be similar in chemical nature to the target analytes
  2. Must elute near the analytes of interest
  3. Must not be present in the actual sample to be analyzed.
  4. Must be available in pure form



- When do we use internal standard calibration?
- Is external calibration enough?

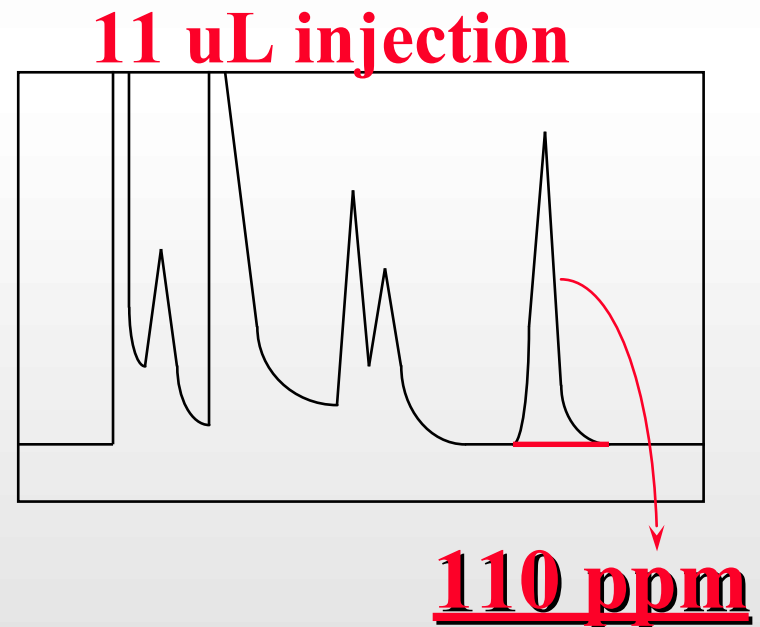
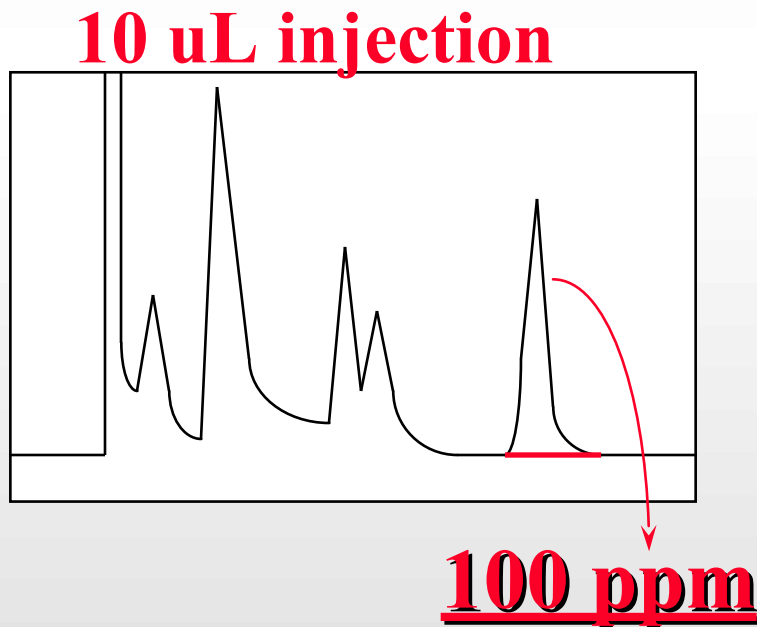
# Advantage of External Standard calibration method

- Only the target compound separation can be focused.



# Disadvantage of External Standard calibration method

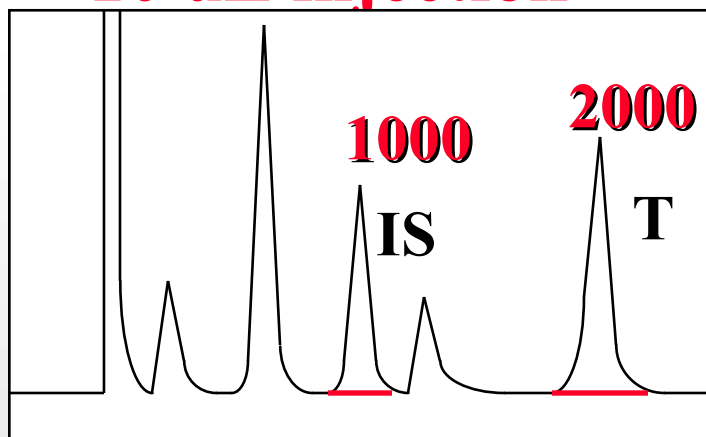
- Injection error will directly influence the quantitative result.



# Advantage of internal standard calibration method

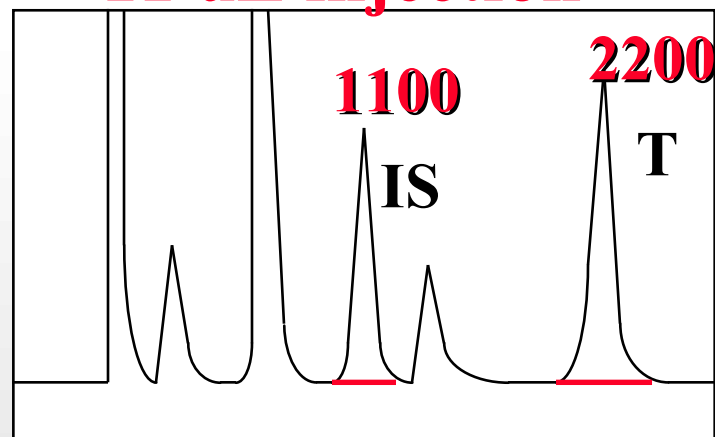
- Injection error can be eliminated.

**10 uL injection**



$$2000 / 1000 = 2$$

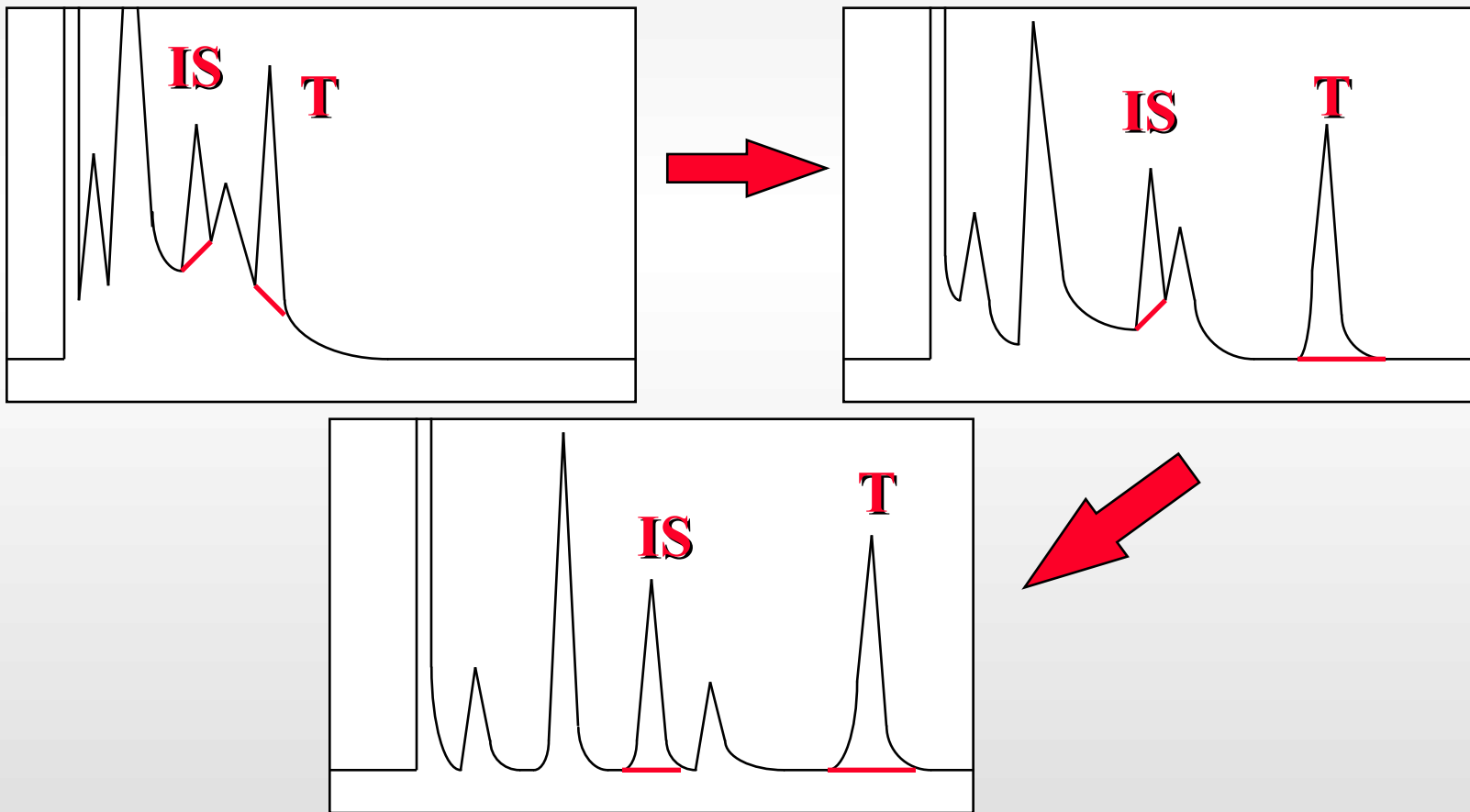
**11 uL injection**



$$2200 / 1100 = 2$$

# Disadvantage of internal standard calibration method

- Separation is slightly difficult.



# Disadvantage of IS Method

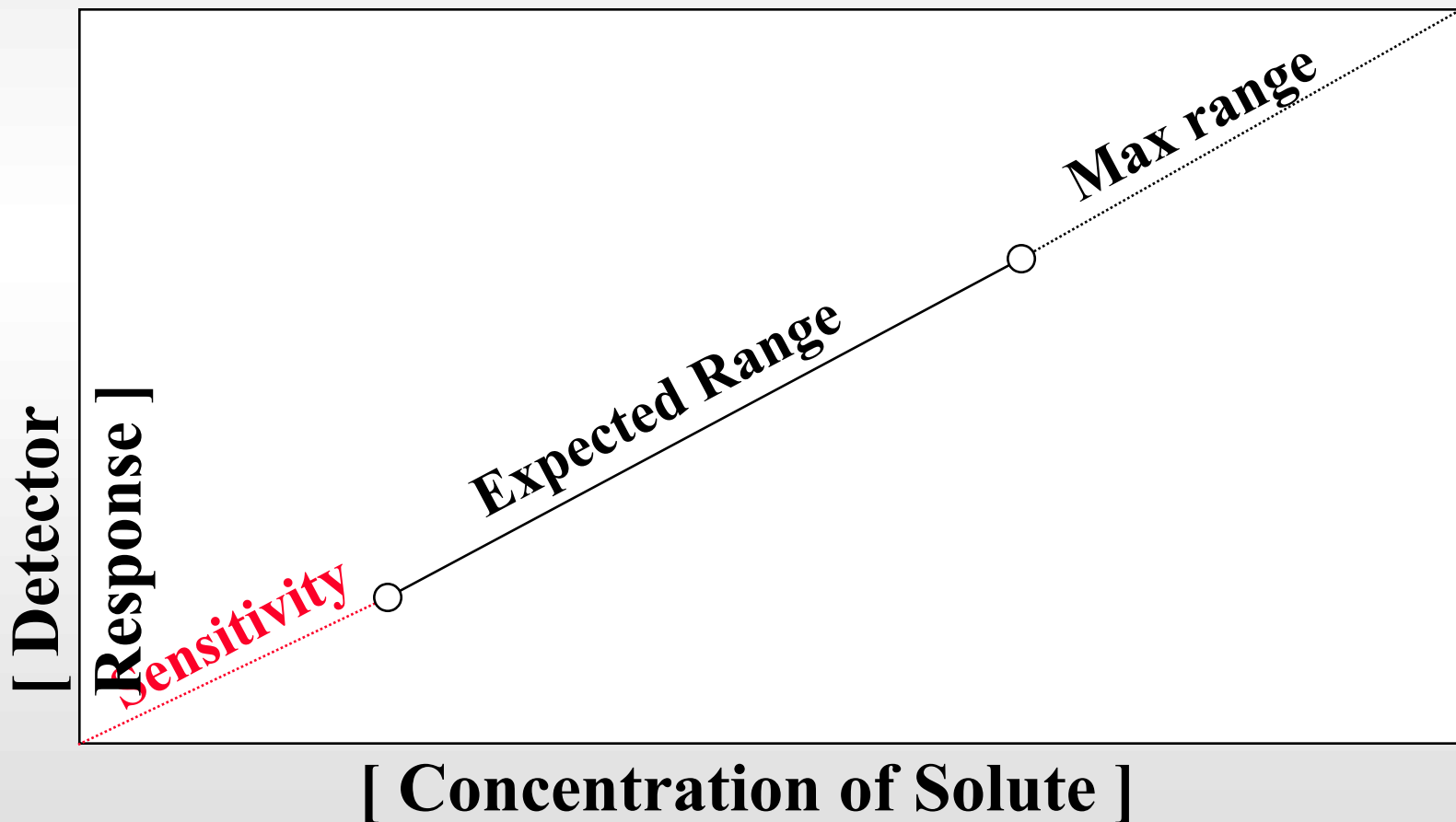
- It is difficult to look for the IS compound.
  - The chemical structure of IS compound should be similar to target compound.
  - IS sample should not exist in the actual sample.

# Calibration Method

- External standard calibration
  - Separation is not difficult
  - Injection error will directly influence the quantitative result
- Internal standard calibration
  - Injection error can be eliminated
  - Recovery in the pretreatment procedure can be estimated
  - Separation is slightly difficult
  - Difficult to look for the IS compound



# Accurate Quantitation - Select Appropriate Working Range



# Preventive Maintenance

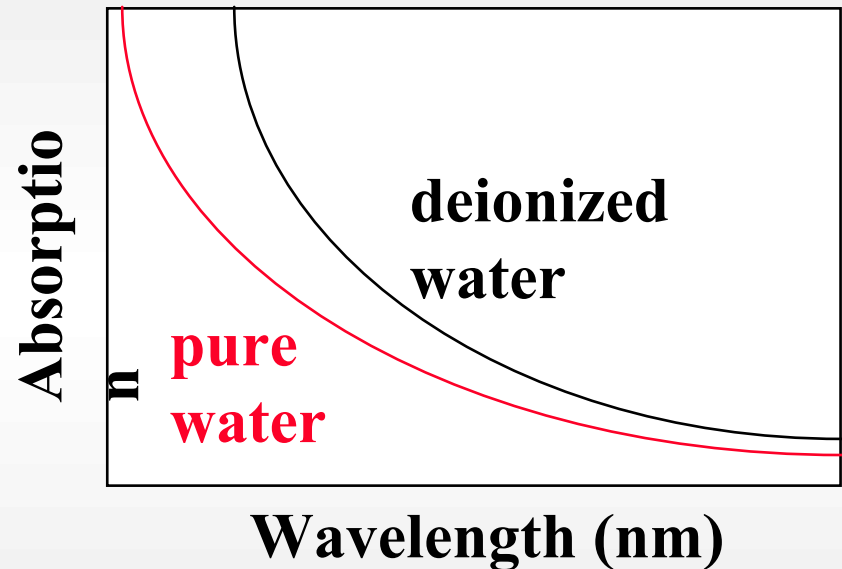


# Common problems in HPLC

- High back pressure
- Poor baseline stability and excessive noise
- Appearance of ghost peaks
- High background absorption of solvent
- Peak tailing
- Pump leakage
- Column failure

# Mobile Phase

- Water
  - Use high purity water
  - Distilled water
  - Deionized water  
(18.2 mega-ohm)
  - Or deionized, distilled water



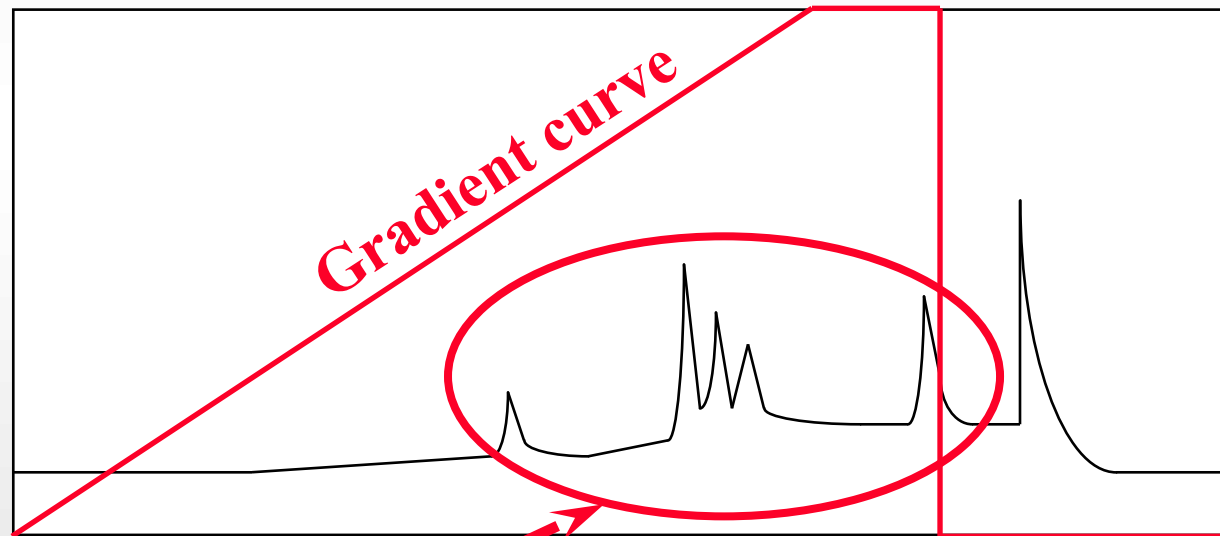
Due to existence of impurity, deionized water may show higher absorption.

# Mobile Phase

- Problem with deionized water is ghost peak appear in gradient elution! (during trace analysis)

**H<sub>2</sub>O / MeOH  
gradient**

**ODS Column**

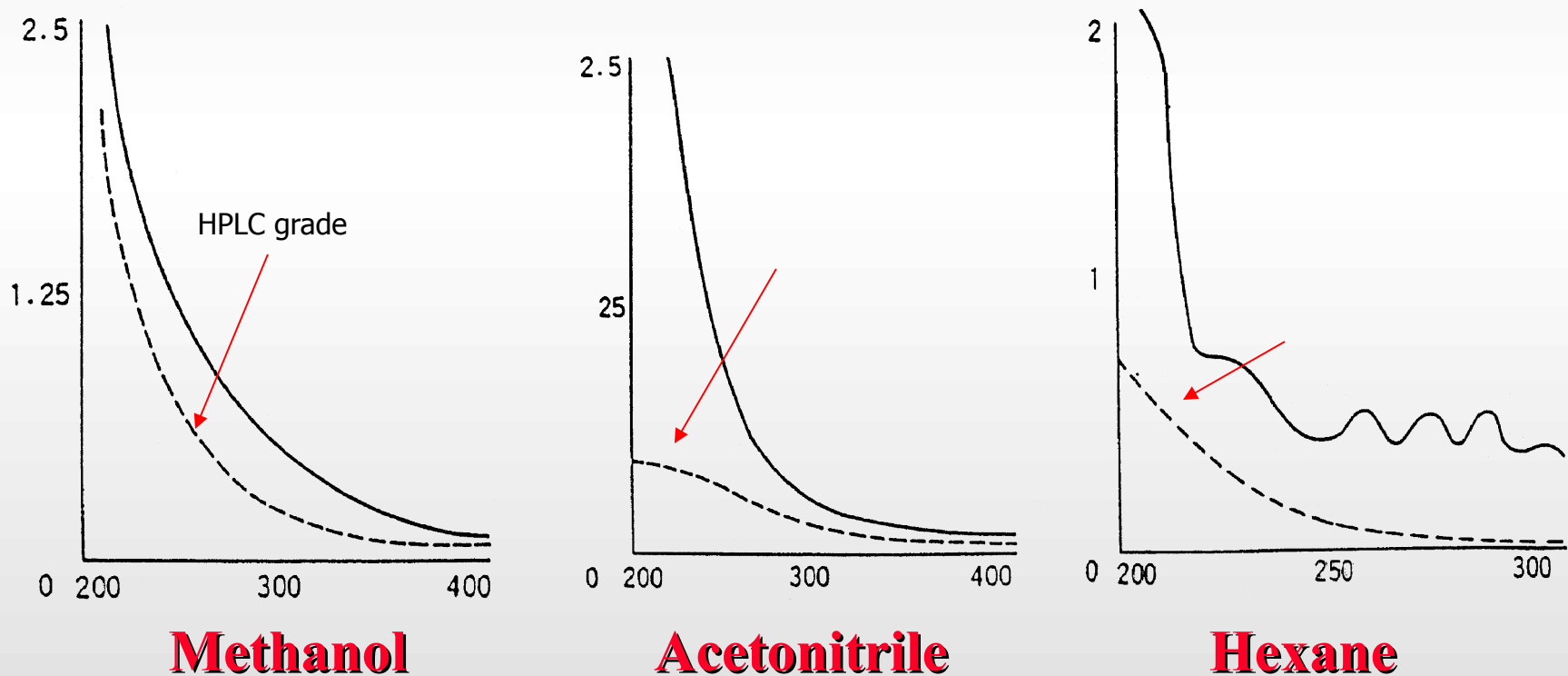


**Ghost Peak**

# Mobile Phase

- Solvents
  - ✓ Use hplc grade

## Difference Between Analytical and HPLC Grade Solvents



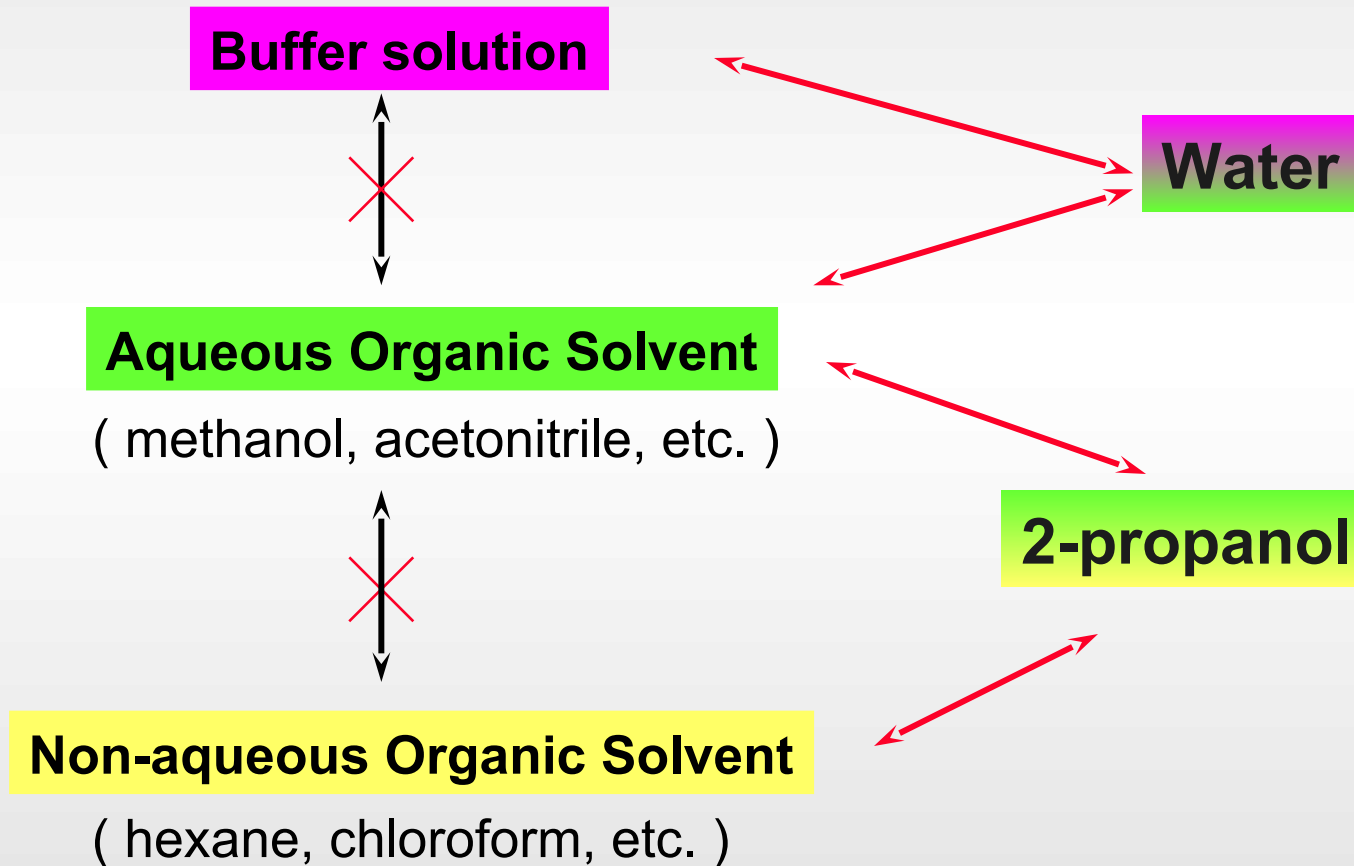
# Mobile Phase

Use solvents with low background absorption

## Cut-off Point for HPLC Solvents

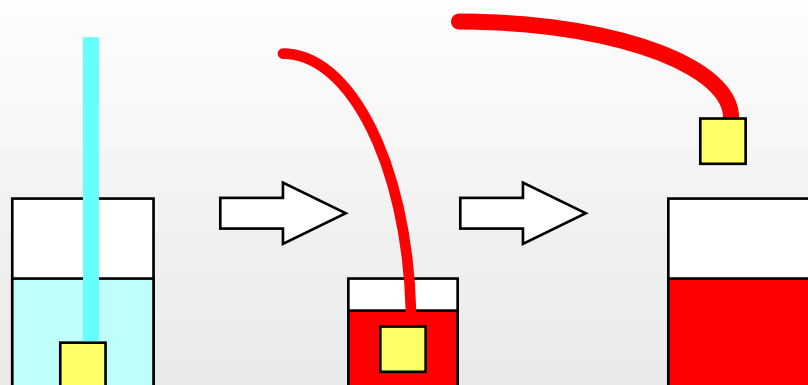
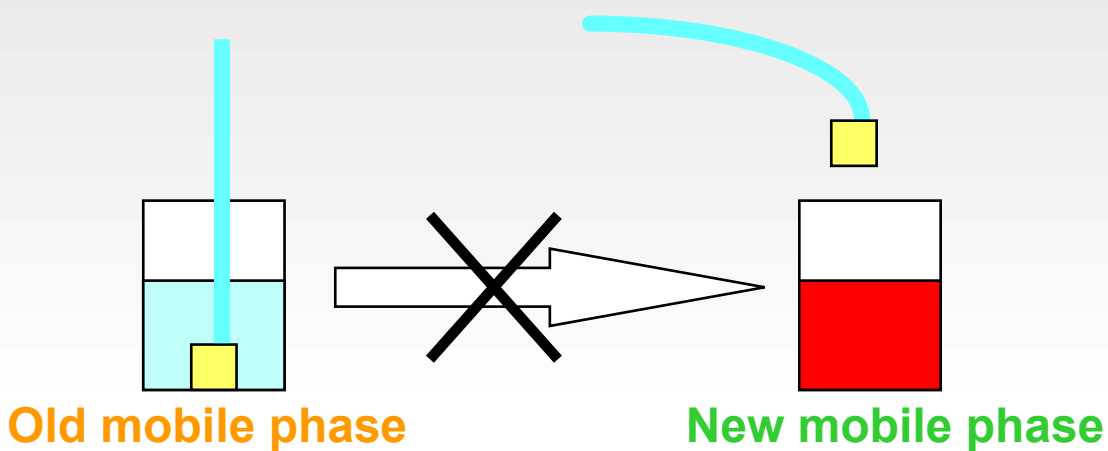
Wavelength	190	195	200	205	210	215	220	230	235	240	245	250	254
Acetonitrile	1.000	0.150		0.070	0.040		0.020						0.010
1-Butanol						1.000	0.500	0.200					0.025
Chloroform											1.000	0.320	0.150
Cyclohexane			1.000	0.880	0.670								0.014
Ethanol				1.000	0.650		0.350						0.040
Ethyl Acetate													1.000
Ethyl Eter							1.000						0.070
Heptane			0.750				0.200						0.014
Hexane		1.000			0.250		0.080						0.014
Methanol				1.000			0.300	0.150					0.025
Methylene Chloride								1.000	0.700	0.200			0.100
Pentane	1.000		0.300		0.100								0.014
2-Propanol				1.000			0.250	0.130					0.025
THF					1.000	0.600		0.300					0.100

# Changing the mobile phase

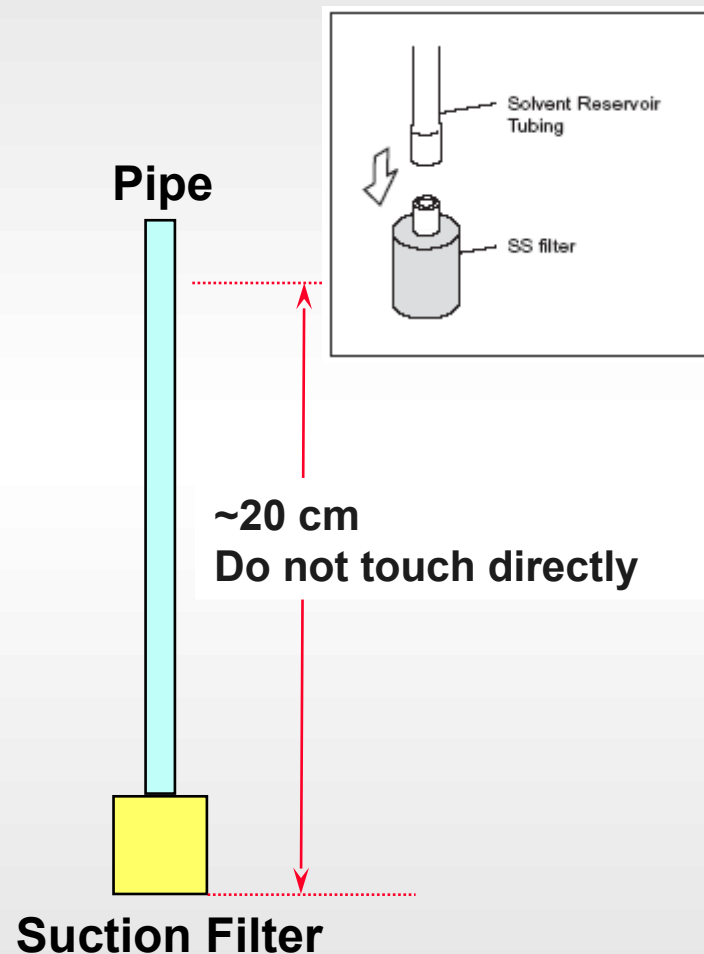




# Changing the mobile phase



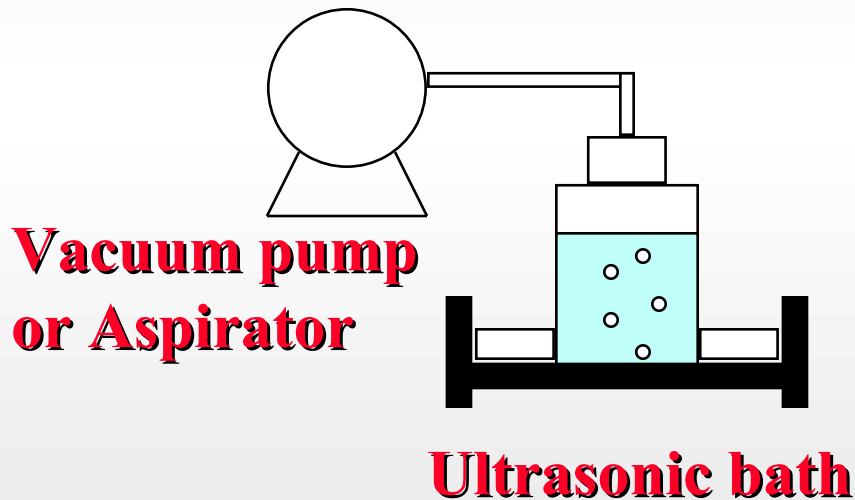
Rinse suction filter  
in fresh mobile phase



# Degassing

Eliminates dissolved gases which can contribute to baseline noise and pump problems.

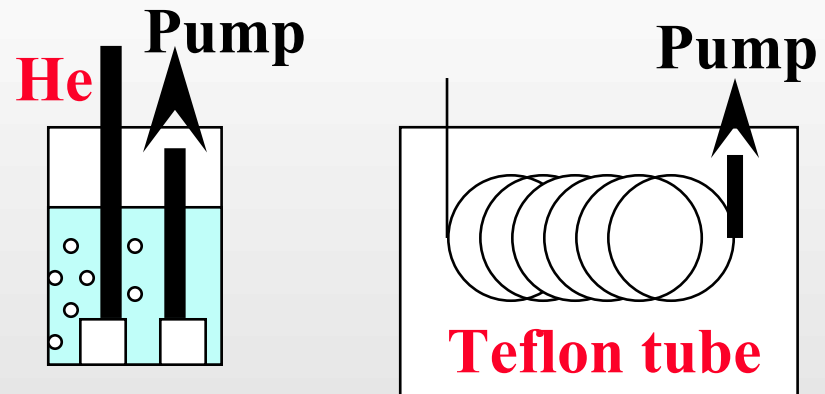
## Off line degassing



## On line degassing

**He purging**

**Vacuum Chamber**



**Vacuum Chamber**

# Reasons for Column Failure

- Plugged Frit or Column Packing
- Adsorbed Sample & Solvent Impurities
- Mechanical Shock, forming Voids
- Chemical Attack of Packing Material

# Plugged Frit or Column Packing

Back-Pressure too High!

- Shortens column lifetime
- Separation sometimes affected
- Caused by particulates in the sample and or mobile phase

# Adsorbed Sample/Solvent Impurities

- Accumulated impurities/contaminants retained in the column can cause peak tailing.

# Prevent Column Clogging

## Avoid Clogging!

- Mobile phase/buffer solutions “must” be filtered using by 0.45 or 0.2  $\mu\text{m}$  membrane filter.
- Samples must be filtered using 0.45 or 0.2  $\mu\text{m}$  membrane filter.
- Don't leave buffer inside the column when not in use



# Remedies for Clogging/Contamination

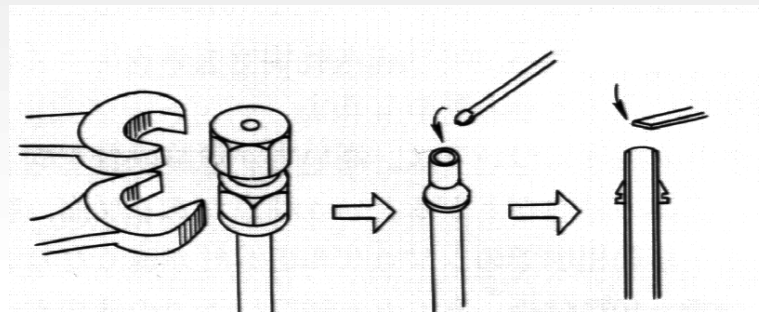
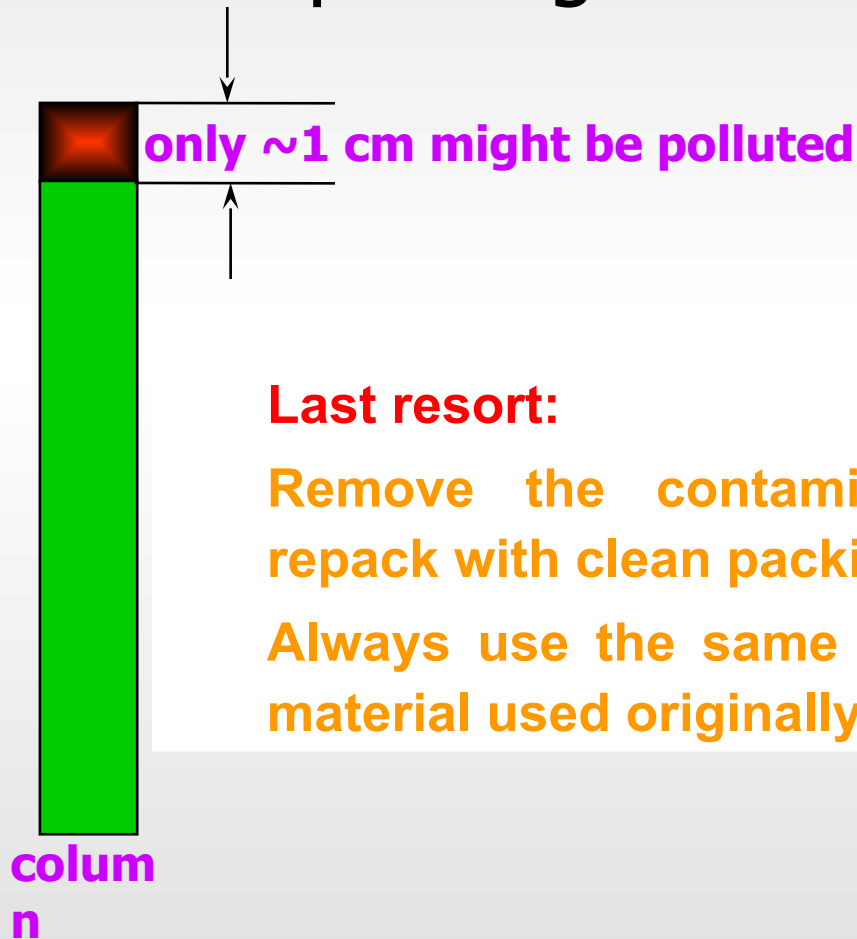
## ■ Washing

For reversed phase column, connect the column in reverse and flow the washing solvents slowly (in sequence).

- 1. Wash with mobile phase without buffer salts
- 2. Wash with methanol or acetonitrile
- 3. Wash with THF or isopropanol
- 4. Wash with hexane
- 5. Wash with THF or isopropanol
- 6. Wash with methanol or acetonitrile

# Remedies for Clogging/Contamination

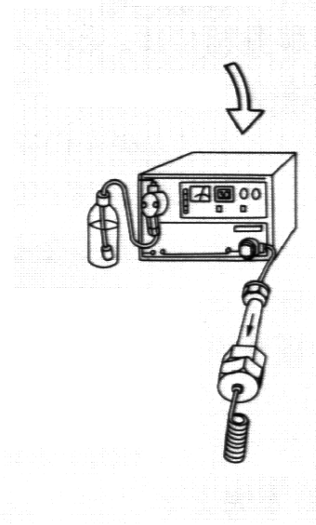
## ■ Repacking



### Last resort:

Remove the contaminated part and repack with clean packing material.

Always use the same size and type of material used originally in the column.





# Causes of Tailing Peaks

- Build-up of garbage on the column inlet
- Extra column effects (dead volume)
- Sample Overload
- Incorrect solvents for the sample
- Secondary retention effects
  - Silanol group
  - Residual heavy metal

# Incorrect solvent for sample

- Avoid selecting a high soluble solvent as sample solvent.

**Methanol** as a sample solvent

**20 uL** Caffeine

A chromatogram showing a single, sharp, and narrow peak for Caffeine. The peak is well-resolved and has a high signal-to-noise ratio, indicating good solubility and stability of the sample in methanol.

**Ethanol** as a sample solvent

**20 uL** Caffeine

A chromatogram showing a peak for Caffeine that is significantly broader and more distorted than the one in methanol. The peak has a shoulder and a tail, indicating poor solubility or instability of the sample in ethanol, which leads to peak broadening and potential loss of resolution.

**Ethanol** as a sample solvent

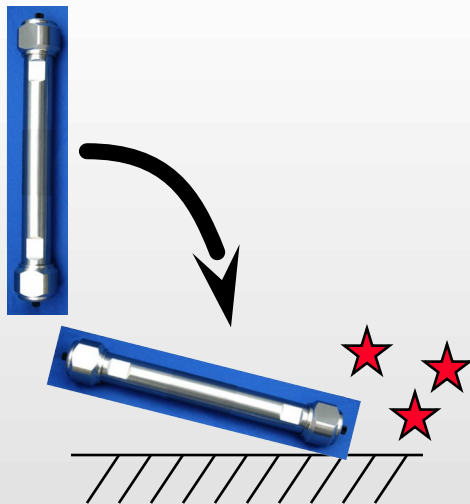
**10 uL** Caffeine

A chromatogram showing a sharp peak for Caffeine in ethanol solvent, but with a smaller injection volume of 10 uL. The peak is much narrower and sharper than the 20 uL ethanol injection, demonstrating that a smaller amount of sample can overcome the solubility issues associated with ethanol.

◆ Better inject small amount of sample.

# Mechanical Shock, forming Voids

- Do not subject the column to shock, such as drop of column or sudden release of pressure. These mechanical shocks will form voids.

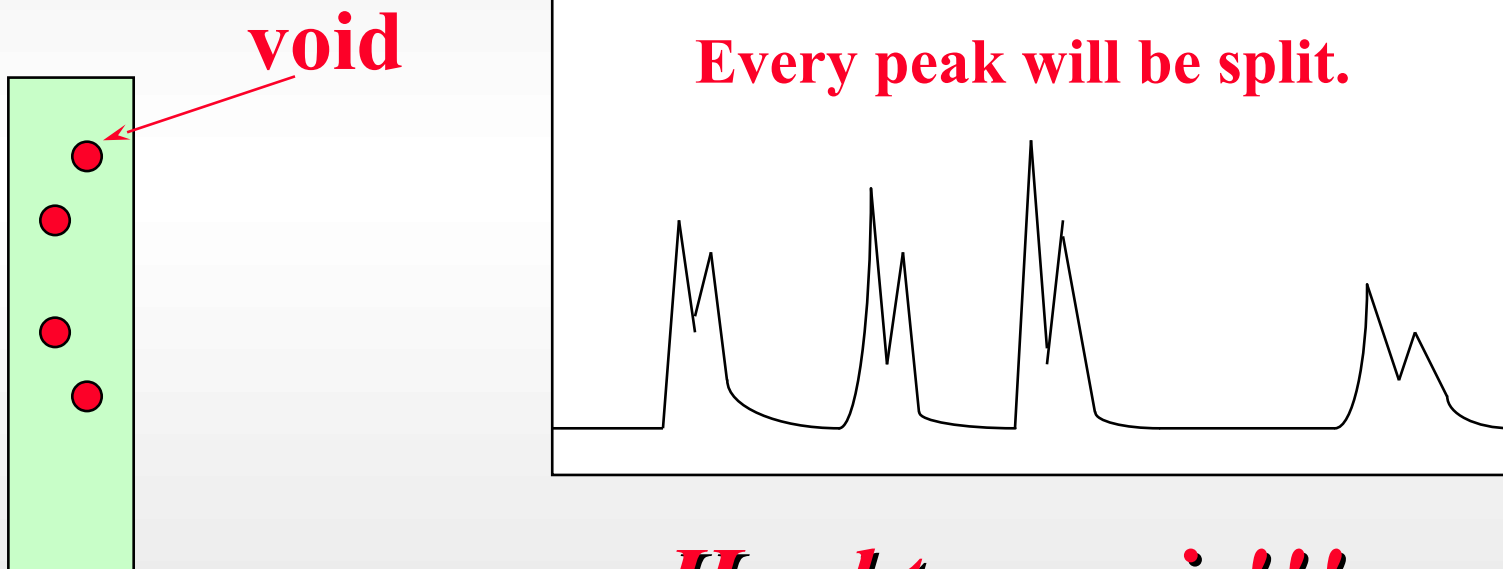


Do not open drain valve during operation



# Voids

- Voids will cause split peaks.



**Hard to repair!!!**

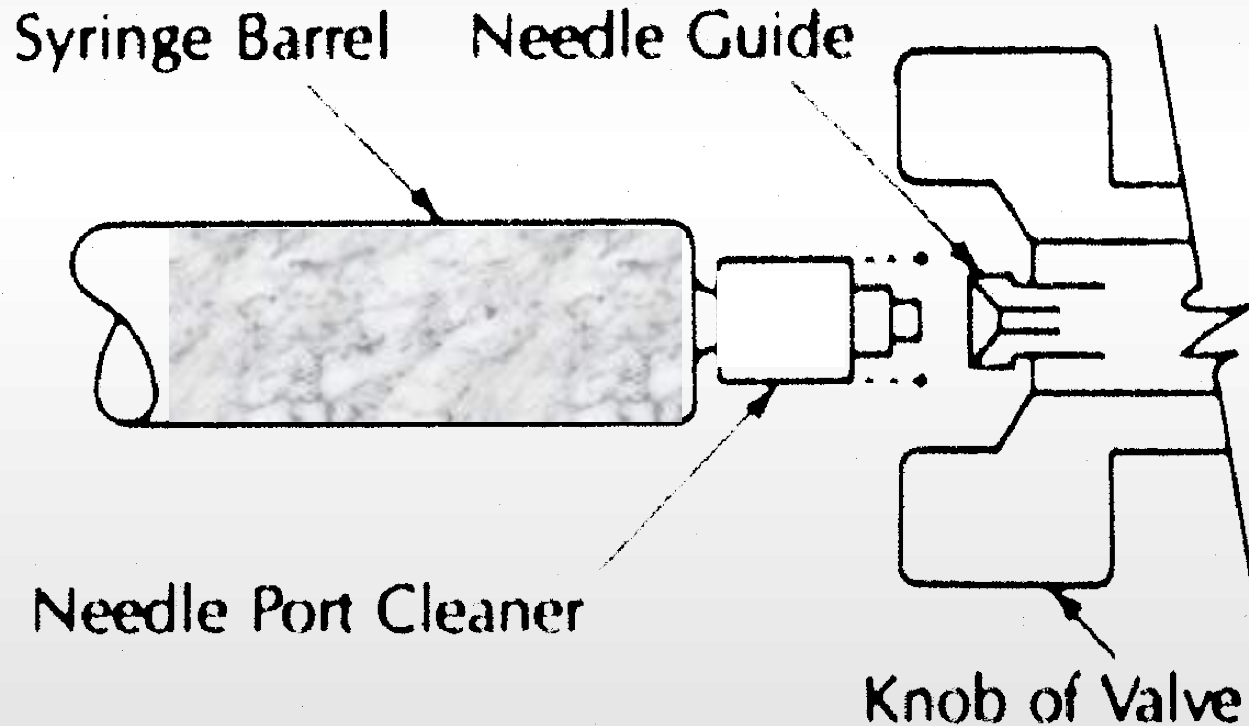
# Precautions for the Packing Material

	Silica gel	Polymer
<b>pH range</b>	<b>2 - 7.5</b>	<b>wider range</b>
<b>Organic solvent</b>	<b>all solvent</b>	<b>limited compatibility</b>
<b>Pressure</b>	<b>less than 250 kgf/cm<sup>2</sup></b>	<b>low pressure</b>
<b>Temperature</b>	<b>better to set at less than 60degC</b>	<b>possible to set at high temperature</b>

**Reminder:** Always read the literature or information sheet that comes with the column!

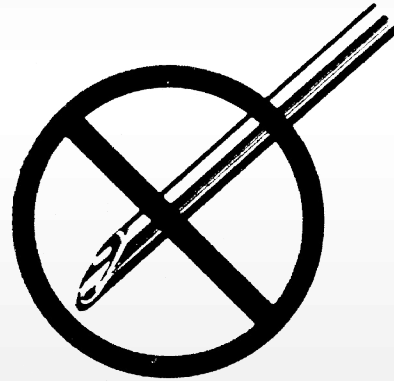
# Washing for Injection Port

- Use of Needle Port Cleaner



# Caution

- Do not use pointed or beveled needle tip.
  - Must use square end type.



- Do not use more than pH 10 solution.
  - Must change rotor seal.

# Maintenance Tips

- Pump
  - check for leaks
  - wash plunger seal after using buffers
- Injector
  - wash injection port after every injection
  - select a suitable washing solvent
- Column
  - do not store in buffer solution
  - Always wash
- Detector
  - check life-time of lamp or electrode